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Running Title: SVR9 regulates chloroplast and leaf development

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Title: Chloroplast translation initiation factors regulate leaf variegation and development

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One-sentence summary

SVR9 and its homologue SVR9L1 encode functionally redundant chloroplast translation initiation factors essential for chloroplast and leaf development in Arabidopsis

Footnotes

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Author contributions

F.Y. conceived and designed research plans; X.L. supervised the experiments; M.Z., S.L. and S.F. performed experiments; M.Z., Y.Q., and J.Z. analyzed the data; L.A. and J.S. provided technical assistance; F.Y. and X.L. wrote the article with contributions from all the authors.

ABSTRACT

Chloroplast development requires the coordinated expressions of nuclear and chloroplast genomes, and both anterograde and retrograde signals exist and work together to facilitate this coordination. We have utilized the *Arabidopsis yellow variegated* (*var2*) mutant as a tool to dissect the genetic regulatory network of chloroplast development. Here, we report the isolation of a new *var2* genetic suppressor locus, *SUPPRESSOR OF VARIEGATION9* (*SVR9*). *SVR9* encodes a chloroplast-localized prokaryotic type translation initiation factor IF3. *svr9-1* mutant can be fully rescued by the *Escherichia coli* IF3 infC, suggesting that SVR9 functions as a *bona fide* IF3 in the chloroplast. Genetic and molecular evidence indicate that SVR9 and its close homologue SVR9-LIKE1 (SVR9L1) are functionally interchangeable and their combined activities are essential for chloroplast development and plant survival. Interestingly, we found that SVR9 and SVR9L1 are also involved in normal leaf development. Abnormalities in leaf anatomy, cotyledon venation patterns and leaf margin development were identified in *svr9-1* and mutants that are homozygous for *svr9-1* and heterozygous for *svr9l1-1* (*svr9-1 svr9l1-1*/+). Meanwhile, as indicated by the auxin response reporter *DR5:GUS*, auxin homeostasis was disturbed in *svr9-1*, *svr9-1 svr9l1-1*/+ and plants treated with inhibitors of chloroplast translation. Genetic analysis established that *SVR9*/*SVR9L1*-mediated leaf margin development is dependent on *CUP-SHAPED COTYLEDON2* activities and is independent of their roles in chloroplast development. Together, our findings provide direct evidence that chloroplast IF3s are essential for chloroplast development and they can also regulate leaf development.

Introduction

One of the most remarkable achievements in nature is the evolution of the semi-autonomous organelles including chloroplasts and mitochondria in eukaryotic cells via endosymbiosis (Dyall et al., 2004; Jensen and Leister, 2014). Current-day chloroplasts are estimated to contain ~3000 proteins, similar to their cyanobacterial ancestors (Leister, 2003). However, during the endosymbiotic process, the vast majority of genes encoding chloroplast proteins has been transferred to the nucleus and became part of the nuclear genome, and consequently higher plants have evolved regulatory pathways that exert control over these nuclear genes for chloroplast proteins (Woodson and Chory, 2008). The fine coordination between the nucleus and the chloroplast has been fulfilled via two-way communications from the nucleus to the chloroplast (anterograde) and also from the chloroplast to the nucleus (retrograde) (Nott et al., 2006; Woodson and Chory, 2012; Chi and Zhang, 2013).

Much of what we know regarding retrograde signaling stemmed from the regulation of the expressions of nuclear genes for chloroplast proteins by the functional and developmental states of the chloroplast. In wild type *Arabidopsis* seedlings, carotenoid biosynthesis inhibitor norflurazon (NF) can induce photobleaching of the chloroplast and trigger the massive down regulation of nuclear genes for chloroplast proteins such as *CHLOROPHYLL A/B BINDING PROTEIN* (*CAB*) (Nott et al., 2006; Chi and Zhang, 2013). Taking advantage of this NF-triggered retrograde response, a series of *genomes uncoupled* (*gun*) mutants that retained *CAB* expressions upon NF treatment have been identified (Susek et al., 1993; Koussevitzky et al., 2007). Based on the *GUN* and other work, the chlorophyll biosynthetic precursor, Mg-protoporphyrin IX (Mg-protoIX), has been shown as one molecule that potentially serves as the signal molecule in the NF-triggered retrograde signaling, although there are reports arguing against this notion (Mochizuki et al., 2008; Moulin et al., 2008). AP2 type transcription factor ABA INSENSITIVE4 (ABI4) has been identified as a nuclear transcription repressor that executes retrograde signaling downstream of chloroplast GUNs (Koussevitzky et al., 2007). Additional retrograde signals and signaling components have also been identified. A screen with low concentration of NF and light intensity has identified additional *happy on norflurazon* (*hon*) mutants (Saini et al., 2011). A chloroplast envelope anchored plant homeodomain transcription factor PTM may be involved in signal transduction from the plastid to the nucleus and PTM was shown to be able to bind the promoter of *ABI4* and activate its expression (Sun et al., 2011). The physiological states of the chloroplast can also trigger signaling to the nucleus (Wilson et al., 2009; Leister, 2012). For example, the redox state of the photosynthetic electron transfer chain has long been thought as a generator of chloroplast-derived signals (Pesaresi et al., 2009; Kindgren et al., 2012). Reactive oxygen species such as singlet oxygen generated from chloroplasts are also able to trigger nuclear gene expression responses, via an EXECUTER1 and 2-mediated pathway

(Wagner et al., 2004; Lee et al., 2007; Galvez-Valdivieso and Mullineaux, 2010; Ramel et al., 2012). Recently, a plastid isoprenoid biosynthetic intermediate, methylerythritol phosphate (MEcPP), was shown to be a new retrograde signal and able to trigger the expression of stress related nuclear genes such as *HYDROPEROXIDE LYASE* (*HPL*) (Xiao et al., 2012). Plastid gene expression (PGE) has also been proposed as a source of retrograde signals (Gray et al., 2003). Chloroplast translation inhibitors such as lincomycin could trigger the down-regulation of photosynthesis associated nuclear genes, which may be part of the broader PGE-mediated retrograde pathway (Pesaresi et al., 2006).

Besides these retrograde signaling pathways that often use the expressions of nuclear genes for chloroplast proteins as the readout, there is also evidence suggesting other modes of retrograde regulation, for example, the influence on overall plant growth and development by the functional and developmental states of the chloroplast (Hricová et al., 2006; Fleischmann et al., 2011; Tiller and Bock; 2014). In the *Arabidopsis thaliana immutans* mutant and the tomato *ghost* mutant, both defective in the plastid alternative oxidase and displaying distinctive green-white leaf variegation phenotypes, there are conspicuous leaf mesophyll cell developmental defects in white leaf sectors (Josse et al., 2000; Aluru et al., 2001). Some *Arabidopsis* albino mutants such as *pale cress* and *cla1* were also shown to have leaves that lack the characteristic palisade tissue (Reiter et al., 1994; Mandel et al., 1996). In contrast to the rapid progressing of the overall field of plastid retrograde signaling, our understanding of the regulation of plant and leaf development by the developmental and functional states of the chloroplast has remained descriptive.

We have used the *Arabidopsis* leaf variegation mutant *var2* as a tool to probe the genetic regulation of chloroplast development (Liu et al., 2010a; 2013; Qi et al., 2015). In this work, we report the identification of a *var2* genetic suppressor gene *SUPPRESSOR OF VARIEGATION9* (*SVR9*). Molecular cloning and functional complementation tests established that SVR9 and its homologue SVR9-Like1 (SVR9L1) act redundantly as chloroplast translation initiation factor IF3. Down-regulation of chloroplast IF3s' activities not only caused chloroplast development defects, but also led to a series of leaf developmental abnormalities including more serrated leaf margin, disorganized mesophyll cells and altered cotyledon venation patterns. We further demonstrated that auxin homeostasis is disturbed in mutants defective in chloroplast IF3s or in plants treated with chloroplast translation inhibitors. Genetic evidence suggests that the NAC family transcription factor CUP-SHAPED COTYLEDON2 (CUC2) is involved in the chloroplast IF3-mediated leaf margin development (Nikovics et al., 2006). Our results suggest that chloroplast translation initiation factors SVR9 and SVR9L1 are required not only for chloroplast development and *var2*-mediated leaf variegation, but also are involved in the coordination of leaf and chloroplast development.

Results

The isolation and cloning of an extragenic *var2* suppressor locus *SVR9*

We have isolated and characterized genetic suppressors of the *var2* leaf variegation mutant through an activation tagging screen in the *var2-5* mutant background (Liu et al., 2010a; Putarjunan et al., 2013). Here we report the isolation of a new *var2-5* suppressor mutant designated *092-004* (Fig. 1A). Following our naming system, the suppressor locus in *092-004* was named *SUPPRESSOR OF VARIEGATION9* (*SVR9*), and the genotype of *092-004* was *var2-5 svr9-1* and the suppressor single mutant was *svr9-1*. Phenotypically, *svr9-1* showed a distinctive virescent phenotype, i.e., young and emerging leaf tissues displayed pronounced chlorosis while these yellow tissues gradually turned green as mutant leaves matured (Fig. 1A). Consistent with the virescent appearance, chlorophylls accumulated at lower levels in young and emerging tissues, but increased in mature tissues in *svr9-1* and *092-004* (Fig. 1B). The suppression of *var2* leaf variegation by *svr9-1* was not allele specific as *svr9-1* can suppress a stronger *var2* allele, *var2-4* (Fig. 1C and 1D).

Initial co-segregation analyses judged by herbicide Basta resistance indicated that *svr9-1* was likely linked with T-DNA, but we were not able to identify the T-DNA insertion site via various techniques. We thus utilized a map-based cloning procedure to identify *SVR9* (Fig. 2). Bulked segregant analysis and subsequent fine mapping narrowed *SVR9* to a ~415 kb region on chromosome 2 (Fig. 2A). Based on the *svr9-1* phenotype, we reasoned that *SVR9* likely codes for a chloroplast-localized protein. DNA sequences of potential nuclear genes for chloroplast proteins in the interval were examined. Surprisingly, PCRs failed to amplify genomic regions around intron 4 of At2g24060 in *svr9-1*, suggesting At2g24060 genomic sequences may be altered in *svr9-1* (Fig. 2B). Consistent with an abnormality at genomic DNA level, At2g24060 transcripts were not detected in *svr9-1* (Fig. 2C). These results indicated that At2g24060 is disrupted in *svr9-1*.

SVR9 encodes a chloroplast translation initiation factor IF3

To confirm that the virescent phenotype of *svr9-1* and the suppression of *var2-5* were indeed due to a potential disruption of At2g24060, we carried out complementation tests. Independent transgenic lines over-expressing At2g24060 under the 35S promoter were able to reverse the virescent phenotype of *svr9-1* and restore the leaf variegation phenotype in *092-004*, thus confirming that At2g24060 is *SVR9* (Fig. 3A).

SVR9/At2g24060 was annotated to encode a putative chloroplast protein of 312 amino acids. SVR9 shows high homologies with translation initiation factor 3 (IF3) proteins in prokaryotes (Supplemental Fig. S1). To test whether the virescent phenotype of *svr9-1* was indeed caused by the lack of a chloroplast IF3, we carried out functional complementation

tests using the *Escherichia coli* IF3 protein infC (Sacerdot et al., 1982). First, we established that the putative chloroplast transit peptide of SVR9 (cTP_{SVR9}, N-terminal 1-71 amino acid residues of SVR9) was sufficient to guide GFP into the chloroplast (Supplemental Fig. S2). Then a binary vector containing a chimeric gene with the cTP_{SVR9} fused at the N-terminus of *E. coli* infC under the control of the 35S promoter was generated (*P_{35S}:cTP_{SVR9}-infC*) and transformed into *svr9-1*. Independent transgenic plants showed green leaf coloration, indicating that cTP_{SVR9}-infC can functionally replace SVR9 (Fig. 3B). Moreover, when *P_{35S}:cTP_{SVR9}-infC* was introduced into *092-004*, transgenic plants showed leaf variegation phenotype resembled that of *var2-5*, suggesting also a functional complementation (Fig. 3B). These results indicate that *E. coli* infC can substitute for SVR9 and the phenotype of *svr9-1* and the suppression of *var2-5* leaf variegation by *svr9-1* were caused by the lack of a chloroplast translation initiation factor IF3.

Phylogenetic analysis of SVR9-like proteins in various photosynthetic organisms showed that higher plant chloroplasts often contain more than one IF3 coding genes (Supplemental Fig. S3). In *Arabidopsis*, two other genes, At4g30690 and At1g34360, also potentially code for prokaryotic IF3-like proteins (Fig. 4A; Nesbit et al., 2015). Of the three genes, *SVR9* and At4g30690 code for putative chloroplast proteins and share the same gene structures, and ~78% identities in amino acid sequences (Fig. 4A; Supplemental Figs. S1 and S3). We thus named At4g30690 as *SVR9-LIKE1* (*SVR9L1*). In contrast, At1g34360 codes for a mitochondrial protein and its gene structure was different from that of *SVR9* or At4g30690 (Fig. 4A; Nesbit et al., 2015). To confirm the chloroplast localizations of SVR9 and SVR9L1, constructs expressing C-terminal GFP fusion proteins SVR9-GFP and SVR9L1-GFP were generated and used to transform *Arabidopsis* wild type leaf protoplasts. When GFP alone is expressed, green fluorescent signals were mainly detected in the cytosol (Fig. 4B). On the other hand, protoplasts expressing SVR9-GFP or SVR9L1-GFP showed signals that clearly aligned with chlorophyll autofluorescence, confirming the chloroplast localization of both proteins (Fig. 4B). Taken together, our results demonstrate that *SVR9* and *SVR9L1* encode the chloroplast translation initiation factor IF3 proteins.

Lastly, we examined the tissue expression pattern of *SVR9*. A ~1.9 kb promoter region upstream of *SVR9* start codon was transcriptionally fused with the *GUS* gene and introduced into wild type *Arabidopsis*. Histochemical GUS staining was performed in *P_{SVR9}:GUS* lines at various developmental stages. In general, strong GUS activities were detected in photosynthetic tissues while GUS levels were below the detection limit in roots (Fig. 4C). In addition, *SVR9* showed stronger expressions in young, newly emerged leaves as compared to the gradually reducing expressions in older leaves (Fig. 4C and 4D). These observations suggest that the expression of *SVR9* is regulated by leaf developmental stages and the need

for SVR9 activities is probably higher in actively growing young tissues, consistent with the virescent mutant phenotypes.

Genetic interactions between *svr9-2*, *svr9l1-1* and *var2*

To further dissect the functions of *SVR9* and *SVR9L1*, and their interactions with *var2*, we sought for additional mutant alleles of these two genes. For *SVR9*, we obtained a SAIL T-DNA insertion line SAIL_172_F02 and confirmed that the T-DNA was located in the last exon of *SVR9*, 175 bp upstream of the stop codon, and this allele was named *svr9-2* (Supplemental Fig. S4). For *SVR9L1*, we obtained a SALK T-DNA insertion line SALK_140431C, which harbored a T-DNA in the 6th intron, and this mutant allele was named *svr9l1-1* (Supplemental Fig. S4). In contrast to the virescent phenotype of *svr9-1*, *svr9-2* showed an overall wild type-like phenotype and reduced accumulation of *SVR9* transcripts, suggesting that it is a weaker allele of *SVR9* (Supplemental Fig. S4). Similarly, *svr9l1-1* mutants were also indistinguishable from wild type (Supplemental Fig. S4). Double mutants of *svr9-2* and *svr9l1-1* also resembled wild type (Supplemental Fig. S4).

We next examined the genetic interactions between *svr9-2*, *svr9l1-1* and two *var2* mutant alleles, *var2-4* and *var2-5* (Supplemental Fig. S5). To evaluate the extent of leaf variegation in a quantitative manner, images of the first true leaves from each genotype were converted to greyscale and distributions of the pixel intensity values of these images were generated and compared. Green sectors of a leaf will have a lower pixel intensity value while white sectors contain pixels with higher intensity values. Uniform leaf color will give rise to a distribution with a relatively narrow peak. In contrast, variegated leaf images have broader peaks and contain more pixels of higher intensity values. Analysis of wild type, *var2-5* and *var2-4* leaf images showed that they each display a distinctive pattern of pixel intensity histogram that correlates well with the degree of variegation (Fig. 5A and 5E). On the other hand, *svr9l1-1*, *svr9-2* and *svr9l1-1 svr9-2* leaves are not only visually indistinguishable from wild type but also have pixel intensity histograms similar to that of wild type (Fig. 5B and 5F). Double mutants of *svr9l1-1* with *var2-5* or *var2-4* showed leaf variegation that resembled *var2-5* and *var2-4*, respectively, suggesting that *svr9l1-1* alone does not significantly alter *var2* leaf variegation (Fig. 5C and 5D). In contrast, double mutants of *svr9-2* with *var2-5* or *var2-4* showed reduced leaf variegation than *var2-5* and *var2-4*, respectively, suggesting that a weaker allele of *SVR9* can partially suppress *var2* leaf variegation (Fig. 5C and 5D). The stronger effects of *svr9-2* compared to *svr9l1-1* in suppressing *var2* variegation were also clearly reflected by the shifts observed in the histograms of *svr9-2 var2* and *svr9l1-1 var2* double mutant leaf images (Fig. 5G and 5H). Interestingly, leaf variegation was almost absent in *svr9l1-1 svr9-2 var2* triple mutants (Fig. 5C-5D, 5G-5H). The fact that *var2* leaf variegation was suppressed to a stronger degree in the *svr9l1-1 svr9-2* background than in the

svr9-2 mutant alone suggested an additive effect of *svr9l1-1* and *svr9-2* on suppressing *var2* leaf variegation. Together, these data suggest that despite their overall wild type appearances, *svr9l1-1*, *svr9-2*, and *svr9l1-1 svr9-2* mutants possess molecular phenotypes, and these phenotypes can modify *var2* leaf variegation.

Chloroplast translation initiation factor IF3 activities are essential for plant development

Given the high homology between *SVR9* and *SVR9L1*, we next tested the functional relationship between these two genes. As *svr9-2 svr9l1-1* double mutants resembled wild type, we sought to obtain *svr9-1 svr9l1-1* double mutants. However, extensive genotyping did not identify double mutants homozygous for both *svr9-1* and *svr9l1-1*. We did recover mutant plants that were homozygous for *svr9-1* but heterozygous for *svr9l1-1* (referred to as *svr9-1 svr9l1-1*/+ hereafter), which showed a stronger virescent phenotype and smaller stature than *svr9-1* alone (Fig. 6A). Genotyping the selfed progeny of *svr9-1 svr9l1-1*/+ did not yield the expected double mutants either. In the siliques of *svr9-1 svr9l1-1*/+ plants, we observed an increased occurrence of abortive seeds (Fig. 6B). In addition, we generated transgenic plants in *svr9-1* background that expressed *SVR9L1* under the control of the 35S promoter. The over-expression of *SVR9L1* effectively rescued the virescent phenotype of *svr9-1* (Fig. 6C). These data suggest that: 1) *SVR9* and *SVR9L1* are functionally redundant; 2) *svr9-1 svr9l1-1* is likely lethal and 3) chloroplast IF3 activities are essential for chloroplast and plant development.

Compromised chloroplast translation initiation factor IF3 activities lead to leaf developmental defects

During the investigation of mutant alleles of *SVR9* and *SVR9L1* and their genetic interactions, we noticed a number of leaf developmental defects in *svr9-1* and *svr9-1 svr9l1-1*/+. First, leaf margin development was perturbed in these mutants. *svr9-1* displayed a more serrated leaf margin compared to that of wild type (Fig. 7A). Leaf margins of *svr9l1-1* resembled wild type, consistent with its wild type appearance (Fig. 7A). However, *svr9-1 svr9l1-1*/+ plants showed an even more pronounced leaf margin serration compared to either of the single mutant (Fig. 7A). The increased leaf serrations from *svr9-1* to *svr9-1 svr9l1-1*/+ was further validated by the quantitative analysis of leaf shape based on three different parameters, the leaf dissection index ($perimeter^2/4\pi \times$leaf area), the number of teeth/leaf perimeter, and the tooth area/leaf area (Royer et al., 2008; Bilsborough et al., 2011) (Fig. 7B-7D). These analyses suggest that decreased activities of chloroplast IF3s promotes the initiation and the outgrowth of leaf lobes. However, we did not observe major leaf serration changes in *svr8-2*, a null mutant lacking the chloroplast ribosomal protein L24, suggesting

that not all mutants of chloroplast translation display abnormal leaf margin development (Supplemental Fig. S6; Liu et al., 2013).

Second, we examined the leaf anatomy of these mutants. Cross sections of wild type leaves reveal the organized arrangement of palisade and spongy mesophyll (Fig. 7E). In *svr9-1*, palisade mesophyll can still be distinguished but is less organized than that of wild type (Fig. 7E). Leaf mesophyll was more dramatically affected in *svr9-1 svr9l1-1*/+, in which the distinction between palisade and spongy mesophyll became obscure and intercellular air space appeared to be increased (Fig. 7E).

Third, we discovered that leaf vascular development is also altered in *svr9-1* and *svr9-1 svr9l1-1*/+ mutants, as indicated by the cotyledon venation patterns. We used the number of closed areoles in mature cotyledons as an indicator of leaf vascular development (Sieburth, 1999). In wild type, cotyledons with two, three and four areoles were common. In *svr9-1*, although the predominant portions of cotyledons show two or three areoles, cotyledons with only one areole were identified in *svr9-1* (Table 1; Fig. 7F). The defects were more conspicuous in *svr9-1 svr9l1-1*/+. The percentage of three or four areoles were drastically decreased in *svr9-1 svr9l1-1*/+ (Table 1; Fig. 7F). Moreover, cotyledons with no closed areoles can be identified in *svr9-1 svr9l1-1*/+ (Table 1; Fig. 7F). Abnormal leaf vascular development is not generally associated with defects of chloroplast development as we did not observe major cotyledon venation changes in an albino mutant *svr4-2* we reported earlier (Supplemental Table S1; Yu et al., 2011).

Taken together, our data show that the down-regulated chloroplast translation initiation factor IF3 activities alter leaf margin, mesophyll and vascular development.

Chloroplast translation defects alter auxin homeostasis

Phytohormone auxin has been implicated in the regulation of leaf margin and vascular development (Scarpella et al., 2006; Bilsborough et al., 2011). Given the margin and vascular defects of *svr9-1* and *svr9-1 svr9l1-1*/+, we explored whether auxin homeostasis was altered in the mutants. To monitor auxin homeostasis, we crossed an auxin reporter line *DR5:GUS* into *svr9-1* and *svr9-1 svr9l1-1*/+ mutant backgrounds, respectively. GUS staining of *DR5:GUS* reporter line seedlings revealed auxin maxima at cotyledon tips, consistent with previous reports (Ulmasov et al., 1997). In contrast, stronger and more expanded DR5:GUS signals were observed in *svr9-1* background compared to that of the *DR5:GUS* reference plants (Fig. 8A and 8B). Clear signals can be observed along the leaf margin in *svr9-1 DR5:GUS* plants (Fig. 8B). A stronger increase in the DR5:GUS staining was observed in the *svr9-1 svr9l1-1*/+ background (Fig. 8C). If the mutations of chloroplast IF3s can lead to abnormal auxin homeostasis, one would expect that the inhibition of chloroplast translation might also cause similar alterations in auxin homeostasis. To test this hypothesis, we treated

DR5:GUS reporter plants with chemical inhibitors of prokaryotic translation, chloramphenicol and spectinomycin, respectively. Compared to plants subjected to the mock treatments, both chloramphenicol and spectinomycin treatments led to stronger and more diffused distributions of auxin signals (Fig. 8D-8F). These data suggest that disturbances in chloroplast translation induced by mutations or pharmacological treatments can affect auxin homeostasis.

We next tested whether auxin homeostasis can impact the development of leaf margins of *svr9-1* and *svr9-1 svr9l1-1*/+. Mutant plants were treated with auxin transport inhibitor 2,3,5-triiodobenzoic acid (TIBA) or *N*-1-naphthylphthalamic acid (NPA). Compared with mock treatments, *svr9-1* and *svr9-1 svr9l1-1*/+ lost the serrated leaf margins upon TIBA or NPA treatments (Fig. 9A and 9B). Consistent with the visual observations, statistical analysis showed that both TIBA and NPA treatments greatly reduced the leaf dissection index of *svr9-1* and *svr9-1 svr9l1-1*/+ (Fig. 9C and 9D). These results indicate that artificially altering auxin homeostasis by drugs could modulate *svr9-1* and *svr9-1 svr9l1-1*/+ mutant leaf margins.

CUC2 is involved in the regulation of leaf margin development mediated by chloroplast translation initiation factor IF3s

It is well established that NAC family transcription factor CUP-SHAPED COTYLEDON2 (CUC2) is a central regulator in auxin-mediated leaf margin development (Nikovics et al., 2006; Kawamura et al., 2010; Bilsborough et al., 2011). To dissect genetically the regulation of leaf margin by chloroplast IF3s, we investigated the genetic interactions between *SVR9*, *SVR9L1* and *CUC2*. We obtained a *CUC2* loss-of-function mutant SALK_035713 (named *cuc2-101* hereafter) and a gain-of-function mutant *cuc2-1D* (Larue et al., 2009). Consistent with previous reports, *cuc2-101* showed a smooth leaf margin while *cuc2-1D* displayed a more serrated leaf margin (Fig. 10A; Nikovics et al., 2006; Larue et al., 2009). We next crossed *svr9-1* and *svr9-1 svr9l1-1*/+ into the *cuc2-101* and *cuc2-1D* background, respectively. Overall, the presence of *cuc2-101* mutation greatly suppressed the leaf dissection index, the number of teeth/leaf perimeter, and the tooth area/leaf area in *svr9-1* (Fig. 10A-10D). Similar suppression of leaf serrations by *cuc2-101* was also observed in *svr9-1 svr9l1-1*/+, albeit to a less extent (Fig. 10A-10D). On the other hand, the addition of *cuc2-1D* to *svr9-1* increased leaf serration based on the three parameters we monitored (Fig. 10B-10D). In *svr9-1 svr9l1-1*/+ background, introduction of *cuc2-1D* mutation has no obvious impact on the leaf dissection index and the number of teeth/leaf perimeter, but *cuc2-1D* did increase the tooth area/leaf area in *svr9-1 svr9l1-1*/+ (Fig. 10B-10D). This is consistent with previous notion that increased *CUC2* expression promotes leaf marginal serrations mostly by increasing the lobe outgrowth (Kawamura et al., 2010). Our observations suggest that *svr9-1* and *svr9-1 svr9l1-1*/+-mediated leaf margin development involves *CUC2*.

Interestingly, despite the genetic interaction between *CUC2* and *SVR9*/*SVR9L1* on the leaf shape characteristics, neither *cuc2* mutants affects the virescent leaf color phenotype of *svr9-1* or *svr9-1 svr9l1-1*/+ (Fig. 10A; Supplemental Fig. S7). Together the genetic interactions between *SVR9* and *CUC2* suggest that *SVR9* and *SVR9L1*-mediated leaf margin development is dependent on CUC2 activities and is independent of their roles in chloroplast development.

Discussion

Chloroplast translation initiation factor IF3 activities are essential for chloroplast and plant development

We are interested in using *var2* as a tool to gain insight into the genetic regulation of chloroplast development (Liu et al., 2010a). In this study, we isolated a new *var2* genetic suppressor locus termed *SUPPRESSOR OF VARIEGATION9* (*SVR9*). We showed that *SVR9* encodes a chloroplast protein bearing high homology to prokaryotic translation initiation factor IF3. In addition, *svr9-1* mutant phenotypes were fully complemented by a bacterial IF3 directed by the chloroplast transit peptide of SVR9. Taken together, we showed conclusively that *SVR9* codes for a *bona fide* chloroplast translation initiation factor IF3.

In *Arabidopsis thaliana*, there are three genes that are annotated to code for prokaryotic type translation initiation factor IF3 (Nesbit et al., 2015). Besides SVR9, we showed that a close homologue of SVR9, which we named SVR9-LIKE1 (SVR9L1), is also a chloroplast protein, while the third gene product is a mitochondrial protein (Nesbit et al., 2015). This complement of prokaryotic type IF3s is consistent with the endosymbiotic origins of the two organelles. For chloroplast-localized SVR9 and SVR9L1, multiple pieces of evidence suggest that they share redundant functions in chloroplast development. *SVR9* and *SVR9L1* share the same gene structures and high homology at amino acid sequence level (Fig. 4A, Supplemental Fig. S1), suggesting that they may have evolved through gene duplication. Functionally, *svr9-1* can be fully rescued by the over-expression of *SVR9L1* (Fig. 6C). Moreover, genetic interaction studies revealed that the combined activities of SVR9 and SVR9L1 are essential for chloroplast and plant development, as we were not able to recover double homozygous mutants for *svr9-1* and *svr9l1-1*. Based on the severity of *svr9-1* and *svr9l1-1* mutant phenotypes, it is likely that SVR9 contributes more to the overall chloroplast IF3 activities while SVR9L1 plays a relatively minor role, and the genetic relationship between *SVR9* and *SVR9L1* indicates an unequal functional redundancy (Briggs et al., 2006).

Three prokaryotic translation initiation factors, IF1, IF2 and IF3, are employed by the 70S ribosome, and IF3 has been proposed to be responsible for sensing translation initiation regions of mRNAs and ensuring initiation fidelity (Lomakin et al., 2006; Milon et al., 2008). Our results indicate that chloroplast translation initiation factor IF3 activities, contributed by

both SVR9 and SVR9L1 in *Arabidopsis* chloroplasts, are indispensable for chloroplast and plant development. Previous studies have indicated that chloroplast translation initiation factor IF2 activities are also essential for plant development (Miura et al., 2007). These data suggest that the proper initiation of translation in the chloroplast is essential for chloroplast biogenesis.

The suppression of *var2* leaf variegation by *SVR9* and *SVR9L1* mutations

Previously we have proposed a threshold model to explain the mechanisms of leaf variegation mediated by VAR2/AtFtsH2 (Yu et al., 2004; 2005). The core of our model is that the development of functional chloroplasts requires a threshold level of thylakoid FtsH complexes/activities. Moreover, we envisioned that this threshold would be dynamic, and changing functional or developmental status of the chloroplast may necessitate different thresholds. Based on the model, we should be able to identify functional or developmental processes that, when disrupted, may modify the threshold and the need for FtsH activities. To this end, we carried out *var2* genetic suppressor screens. In these genetic suppressors of *var2*, we propose that the suppressor mutations lead to disturbed functional or developmental chloroplast states that lowers the endogenous threshold for VAR2/AtFtsH2. Through both our work with the *SUPPRESSOR OF VARIEGATION* loci and work from other groups, a definitive link between the threshold and some aspects of chloroplast translation has been established (Miura et al., 2007; Yu et al., 2008; Liu et al., 2010b; 2010c; 2013). However, a key aspect of the threshold model, i.e. a multi-point correlation of the changing functional or developmental states of the chloroplast with the corresponding changing needs for FtsH activities, remained hypothetical. In this study, through multiple mutant alleles and mutant combinations of *SVR9* and *SVR9L1*, we were able to create a series of chloroplast developmental states and explore this important aspect of the threshold hypothesis. In wild type background where chloroplast IF3s are at full capacity, VAR2/AtFtsH2 activities are necessary for chloroplast development and the lack of VAR2/AtFtsH2 causes leaf variegation. The *svr9l1-1* mutation slightly reduces IF3 activities, but functional VAR2/AtFtsH2 is still required and leaf variegation is not suppressed. However, in *svr9-2*, the reduction of IF3 activities is greater than in *svr9l1-1*. The state generated by *svr9-2* may lower the threshold need for VAR2/AtFtsH2 and leaf variegation reduces. IF3 activities are further reduced in *svr9l1-1 svr9-2* double mutants, and the mutant background of *svr9l1-1 sv9-2* generated a state that the need for VAR2/AtFtsH2 activities is no longer necessary for chloroplast development, thus a full suppression of leaf variegation. When IF3 activities is further reduced in *svr9-1* and *svr9-1 svr9l1-1*/+ mutants, the lack of IF3 activities itself starts to impair chloroplast development and mutant plants display a clear visual phenotype, and this

functional state bypass the need for VAR2/AtFtsH2, as observed in our previous *var2* suppressor mutants (Liu et al., 2010a; Putarjunan et al., 2013).

Several messages can be gathered from these data. First, our genetic data indicate that full wild type level of IF3 activities are not necessary for chloroplast development under lab conditions, as *svr9l1-1*, *svr9-2*, and *svr9l1-1 svr9-2* resembled wild type. However, a key message from our data is that a superficial wild type appearance does not necessarily mean a lack of phenotypes at molecular level. In this sense, genetic suppressor screens can be a very sensitive and powerful approach to uncover subtle molecular alterations in mutants. Second, the different mutants and mutant combinations in our study generated different chloroplast states where VAR2/AtFtsH2 functions may be partially or completely bypassed, supporting our previous hypothesis that the threshold for FtsH activities can vary.

The interaction between chloroplast and leaf development

The chloroplast is the site for photosynthesis, as well as the venue for the synthesis of numerous metabolic and regulatory molecules such as phytohormones, and the proper functioning of the chloroplast is essential for plant growth and development. It is thus not surprising that the functional or developmental states of the chloroplast can impact processes outside of the chloroplast. Previous studies have established multiple regulatory pathways that coordinate the states of the chloroplast with gene expressions in the nucleus. In addition, it is widely known that plant leaf development is also linked with the states of the chloroplast. However, we lack detailed information regarding the relationship between leaf development and the chloroplast.

In this study, we uncovered a range of leaf developmental phenotypes in *SVR9* and *SVR9L1* mutants that may be linked with altered auxin homeostasis and auxin regulated pathways. First, we discovered that the cotyledon venation patterns were abnormal in *svr9-1* and *svr9-1 svr9l1-1*/+ mutants. Overall, we observed that mutant cotyledons showed a trend of reduced complexity of vascular development as indicated by the numbers of areoles (Fig. 7F; Table 1). It is generally believed that auxin is a central regulator of leaf vascular development, thus the venation pattern changes in the mutants may reflect an altered auxin homeostasis (Sieburth, 1999; Cheng et al., 2007). Second, we observed that leaf margins are more serrated in *svr9-1* and *svr9-1 svr9l1-1*/+ mutants than those of the wild type (Fig. 7A-7D). The induction of leaf serrations are dictated by a feedback loop of auxin maxima established by PIN1 convergence points and CUC2 activities (Bilsborough et al., 2011). The expression of *CUC2* promotes the formation of PIN1 convergence points and in turn the auxin maxima along the leaf margin where *CUC2* expressions are then repressed by auxin (Bilsborough et al., 2011). This feedback regulation of CUC2, PIN1 and auxin level eventually leads to the alternate presence of *CUC2* expressions at leaf indentations and auxin

maxima at leaf protrusions (Bilsborough et al., 2011; Byrne, 2012). Thirdly, we observed alterations in auxin homeostasis in *svr9-1* and *svr9-1 svr9l1-1*/+ mutants as indicated by the changes in *DR5:GUS* expression patterns (Fig. 8A-8C). Changes in auxin homeostasis can also be observed in plants treated with chloroplast translation inhibitors (Fig. 8D-8F). Treatments with auxin transport inhibitors can alter the leaf margin phenotypes of *svr9-1* and *svr9-1 svr9l1-1*/+ mutants (Fig. 9). These data suggest that auxin may play an important role in leaf developmental processes regulated by chloroplast IF3s. Surprisingly, we did not observe altered leaf margins in *svr8-2*, a null mutant of the chloroplast ribosomal protein L24 (Supplemental Fig. S6; Liu et al., 2013). It is possible that when individual components of the chloroplast translation system were mutated, chloroplast translation is affected in such a manner that may or may not trigger the abnormal development of leaf margins. This raises the interesting possibility that specific aspects of chloroplast translation are associated with leaf margin development. Last but not least, our genetic data support the notion that chloroplast IF3s SVR9 and SVR9L1 regulate leaf margin development at least partially via the *CUC2*-mediated pathway since a lack of *CUC2* greatly hampered but not completely abolished the manifestation of leaf margin phenotypes of *svr9-1* and *svr9-1 svr9l1-1*/+ (Fig. 10). The genetic interaction results further suggest that there may be other factors involved in the output of the retrograde signals from the *svr9-1 svr9l1-1*/+ mutant chloroplasts (Fig. 10). Based on our data, it is clear that certain functional states of the chloroplast, as generated by the lack of *SVR9* and *SVR9L1*, are able to regulate leaf development. It is possible that these states of the chloroplast are capable of altering the auxin homeostasis, which in turn regulates CUC2 activities. It is also possible that these states influence auxin and CUC2 separately. Although the precise mechanisms remain to be elucidated, it is tempting to speculate that these functional states may generate retrograde signals that coordinate the states of the chloroplast with leaf development.

Conclusions

Taken together, our data indicate that chloroplast translation initiation factor IF3 activities are essential for plant and chloroplast development. Mutations in the chloroplast translation initiation factor IF3s can suppress the leaf variegation phenotype of *var2* mutants. Moreover, our data indicate that the functional or developmental states of the chloroplast can regulate leaf development, particularly the processes that are associated with auxin. Currently we do not know whether this regulation involves the known retrograde signaling pathways or it represents a new route of signaling, and future research will undoubtedly expand our understanding of the intricate relationship between the chloroplast and the rest of the plant cell.

Materials and methods

Plant materials and growth conditions

All *Arabidopsis thaliana* strains used in this study are of the Columbia-0 background. *svr9-1* was isolated in this study. The T-DNA insertion lines SAIL_172_F02 (*svr9-2*), SALK_140431C (*svr9l1-1*), SALK_035713 (*cuc2-101*) and *cuc2-1D* were obtained from the Arabidopsis Biological Resource Center (ABRC); *svr4-2* and s*vr8-2* have been described (Yu et al., 2011; Liu et al., 2013). Seeds were sown on commercial soil mix (Pindstrup, Denmark) or on half strength Murashige and Skoog (1/2 MS) medium supplemented with 1% sucrose, and stratified for two days at 4ºC before placed in growth rooms. When solid medium was needed, 0.8% agar (w/v) was added to the 1/2 MS medium. *Arabidopsis* plants were maintained at ~22 ºC under continuous illumination (~100 μmol m^{-2} s^{-1}).

Chlorophyll content measurements

Fresh plant tissues were harvested, weighed, and finely ground in liquid N_2. Total chlorophyll was extracted with 95% ethanol. Chlorophyll contents were determined as described (Lichtenthaler, 1987). Chlorophyll measurements of different samples were normalized on a fresh tissue weight basis. Each sample consisted of leaf tissues pooled from 2 to 4 individual plants. Mean chlorophyll content of leaf tissues of each genotype was calculated from three independent pooled leaf samples. Differences in chlorophyll content between WT and mutants were evaluated with *p* values generated by *t*-test.

Map-based cloning

The *SVR9* locus was initially mapped by the bulked segregation analysis using a pool of 95 F2 mutant seedlings from a cross between *svr9-1* and Landsberg *erecta* (Lukowitz et al., 2000). A mapping population consisting of 570 F2 mutant plants was used to further fine map the *svr9-1* mutation. Primers of the molecular markers generated in this study were listed in Supplemental Table S2. Primers used to amplify various At2g24060 genomic fragments were listed in Supplemental Table S2.

Semi-quantitative RT-PCR analysis

Total cellular RNAs were purified using Trizol RNA reagent (Life Technologies, CA, USA) following the manufacturer's instructions and stored at -80 ºC. For semi-quantitative RT-PCR analysis, first strand cDNAs were synthesized from 1 μg DNase I treated total RNAs using a PrimeScript reverse transcription kit (Takara, Japan). The gene-specific primers were listed in Supplementary Table S2.

Functional Complementation of *svr9-1*

All primers used in vector construction were listed in Supplementary Table S2. To complement *svr9-1*, full-length cDNAs of *SVR9*/At2g24060 and *SVR9L1*/At4g30690 were amplified using primers 24060F and 24060R, and 30690F and 30690R, respectively. The amplified fragments were cloned into pBluescript KS+ (pBS) and sequenced before subcloned into a binary vector pBI111L (Yu et al., 2004). The resulting constructs were named P_{35S}*:At2g24060* and P_{35S}*:At4g30690*, respectively.

To express *E. coli* IF3 infC in *Arabidopsis*, the coding sequence of *infC* was amplified from *E. coli* genomic DNA using primers *E. coli* IF3-F and *E. coli* IF3-R and cloned into pBS. The coding sequence of the chloroplast transit peptide (cTP) region of SVR9 (1-71 aa of SVR9) was amplified with cTPF and cTPR and cloned into pBS. After sequencing pBS-infC and pBS-SVR9cTP, the *infC* and *SVR9cTP* were sequentially subcloned into pBI111L to have SVR9cTP fused in frame at N-terminal of infC. The resulting construct was named P_{35S}*:cTP*$_{SVR9}$*-infC*.

Each of the binary vectors was transformed into *Agrobacterium tumefaciens* by electroporation, and the floral dip method was used for *Arabidopsis* transformation (Clough and Bent, 1998). T1 transgenic lines were screened on solid 1/2 MS medium containing 50 mg·L^{-1} kanamycin.

Protoplast transient expression assays

To transiently express C-terminal GFP fusion proteins SVR9cTP-GFP, SVR9-GFP and SVR9L1-GFP in leaf protoplasts, the coding sequences of SVR9cTP, SVR9 and SVR9L1 were cloned into pTF486 to fuse in frame with GFP coding sequences (Yu et al., 2008). Fresh prepared wild type *Arabidopsis* mesophyll protoplasts were transformed with each of the transient expression vectors following the procedures described in Yoo et al. (2007). After transformation, protoplasts were incubated overnight and examined with confocal microcopy using the Apo TIRF 60x Oil objective (N.A. 1.49) (Nikon A1). GFP and chlorophyll autofluorescence signals were monitored through 525/50 and 700/75 emission filters, respectively.

Histochemical GUS staining

To generate *SVR9* promoter-β-glucuronidase (GUS) transcriptional fusion construct, a 1913 bp genomic DNA fragment upstream of the start codon of *SVR9* gene was amplified using primers 24060PF-XbaI2 and 24060PR-BamHI2 and cloned into pCB308 (Xiang *et al.*,1999). The resulting construct was used to transform wild type *Arabidopsis* plant. P_{SVR9}*:GUS* lines were screened based on Basta resistance. GUS activities were assayed in T1 and T2 generations (Jefferson, 1987).

To test the impact of *svr9-1* and *svr9l1-1* mutations on *DR:GUS* expression patterns, *DR5:GUS* reporter gene was introduced to *svr9-1* and *svr9-1svr9l1-1*/+ via genetic crossing. DR5:GUS activities in different genetic backgrounds were assayed in 6-day-old seedlings.

Quantification of leaf variegation

The first true leaves of two-week-old *Arabidopsis* seedlings were hand cut and mounted in water. Images of each individual leaf were captured using a stereoscope (Nikon, SMZ25) equipped with a CCD camera (Nikon, DS-U3). All leaves were photographed with the same settings. To quantify the extent of variegation, leaf images were first converted to grayscale using the ImageJ software. Pixels constituted the leaf blade were selected and intensity values of these pixels were obtained using the NIS-Elements software (Nikon). Frequency was calculated as number of pixels of each intensity value (ranging from 0-255)/number of pixels constituting the leaf blade area. Frequency distributions of pixel intensity values of each leaf image were generated using the GraphPad Prism software. The quantification of leaf variegation was repeated three times. Similar patterns of frequency distributions of pixel intensity values were obtained for each repeat.

Leaf margin analysis

Water mounted individual leaves were photographed using a stereoscope (Nikon, SMZ25) equipped with a CCD camera (Nikon, DS-U3). Three parameters that were commonly used in leaf margin analysis including: the leaf dissection index ($perimeter^2/4\pi \times$leaf area), the number of teeth/leaf perimeter, and the tooth area/leaf area, were used in this study to quantify the differences in leaf margin (Royer et al., 2008; Bilsborough et al., 2011). Leaf blade region (excluding leaf petiole) was used in quantitative analyses. Leaf area and perimeter measurements were performed using the ImageJ software. Graphs were generated by the GraphPad Prism software. Differences between means of each leaf shape parameter were evaluated by *p* values generated by *t*-test.

Analysis of leaf cross sections and cotyledon vein patterns

Basal parts of the first of true leaf of 10-day-old wild type, *svr9-1* and *svr9-1 svr9l1-1*/+ were hand cut and infiltrated with fixation solution (4% (v/v) glutaraldehyde in 0.1 mM sodium phosphate buffer, pH6.8). Fixed tissues were dehydrated and embedded in Technovit 7100 resin (EMS, PA, USA). Semi-thin sections (10 μm) prepared with a microtome (RM2265, Leica) were stained with 1% (v/v) toluidine blue O and observed with a Leica DM5000B microscope equipped with a DFC425C CCD camera.

To observe cotyledon veins, cotyledons of 10-day-old plants were hand cut and decolored in 70% ethanol until the veins became clearly visible. Decolored cotyledons were

618 then photographed with a digital camera mounted on a stereo microscope. Cotyledon vein
619 patterns were categorized based on the number of areoles formed (Sieburth, 1999)
620
621 **Chemical treatments**
622 All chemicals used in this study were purchased from Sigma unless otherwise specified.
623 1-N-naphthylphthalamic acid (NPA) and 2,3,5-Triiodobenzoic acid (TIBA) were dissolved in
624 dimethyl sulfoxide (DMSO). Wild type, *svr9-1* and *svr9-1 svr9l1-1*/+ seeds were germinated
625 and grown on solid 1/2 MS medium supplemented with 1 μM NPA or 1 μM TIBA. Plants
626 grown on solid 1/2 MS medium with equal concentration of DMSO served as controls.
627 Images of the third true leaves in 2-week-old plants were examined.
628 To test *DR5:GUS* expression in response to chloroplast translation inhibitors, *DR5:GUS*
629 lines of different genetic backgrounds were germinated and grown on liquid 1/2 MS medium
630 for 6 days before adding chloramphenicol or spectinomycin to a final concentration of 5 mM.
631 Equal amounts of ethanol were added as the mock treatment. After 24 hours of treatment,
632 seedlings were harvested and assayed for GUS activities. Plant liquid cultures were shaken at
633 120 rpm to keep the seedlings floating.
634
635 **Accession numbers**
636 Sequence data from this article can be found in the The *Arabidopsis* Information Resource
637 (TAIR) or GenBank/EMBL databases under the following accession numbers: SVR9,
638 At2g24060; SVR9L1, At4g30690; VAR2/AtFtsH2, At2g30950; CUC2, At5g53950, *E. coli*
639 infC, NP_416233.1.
640
641 **Supplemental data**
642 **Supplemental Table S1.** Comparison of cotyledon vein patterns of wild type and *svr4-2.*
643 **Supplemental Table S2.** Primers used in this study.
644 **Supplemental Figure S1**. Protein sequence alignments of SVR9/At2g24060,
645 SVR9L1/At4g30690, At1g34360 and *E. coli* infC.
646 **Supplemental Figure S2.** Chloroplast localization of cTP_{SVR9}-GFP.
647 **Supplemental Figure S3.** Phylogenetic analysis of prokaryotic IF3-like proteins from *E. coli*
648 and representative photosynthetic species.
649 **Supplemental Figure S4.** Identification of *svr9-2* and *svr9l1-1*.
650 **Supplemental Figure S5.** Whole plant phenotypes of representative 2-week-old plants of the
651 same genotypes shown in Fig. 5A-5D.
652 **Supplemental Figure S6.** Comparison of leaf margin development in WT, *svr9-1* and *svr8-2*.
653 **Supplemental Figure S7.** Whole plant phenotypes of representative 2-week-old plants of the
654 same genotypes shown in Fig. 10A.

655 **Figure legends**

656 **Figure 1.** Suppression of *var2* leaf variegation by *svr9-1*. A, Representative 15-day-old wild
657 type (WT), *var2-5*, *092-004* (*var2-5 svr9-1*), and *svr9-1* plants. C, Representative 15-day-old
658 WT, *var2-4*, *svr9-1*, and *var2-4 svr9-1* double mutant. B and D, Chlorophyll contents of
659 indicated tissues in plants shown in A and C, respectively. Total: entire rosettes excluding the
660 cotyledons; Peripheral: the first two true leaves; Center: rosettes excluding the cotyledons and
661 the first two true leaves. Chlorophyll measurements were normalized on a fresh tissue weight
662 basis. Data were presented as mean ± standard deviation (s.d.) of three biological replicates.
663 *: $0.01<p<0.05$; **: $p<0.01$.
664
665 **Figure 2.** Cloning of *SVR9*. A, Schematic representation of the positional cloning procedure
666 used to identify *svr9-1* mutation. Primers of the molecular markers designed in this study
667 were listed in Supplemental Table S2. A total of 570 F2 plants (1140 chromosomes) were
668 used in fine mapping. Numbers of recombinants were marked under each molecular marker.
669 The asterisk indicated the position of *SVR9* (At2g24060). In the gene model, boxes represent
670 exons and lines represent introns. 5' and 3' untranslated regions (UTRs) were shaded. Arrows
671 represent the positions of the primers used in B and C. B, PCR analyses using various primer
672 combinations indicated that At2g24060 genomic regions were disrupted in *svr9-1*. C, Semi-
673 quantitative RT-PCR analyses of the accumulations of *SVR9* transcripts in wild type and *svr9-*
674 *1* using indicated primer combinations. Expression of *ACT2* was used as an internal control.
675
676 **Figure 3.** Complementation of *svr9-1* and *092-004*. A, Leaf color phenotypes of two-week-
677 old WT, *svr9-1*, *var2-5*, *092-004* and representative lines overexpressing At2g24060 in *svr9-1*
678 or *092-004* background. B, Leaf color phenotypes of two-week-old WT, *svr9-1*, *var2-5*, *092-*
679 *004* and representative lines overexpressing CTP_{SVR9}-infC in *svr9-1* or *092-004* background.
680
681 **Figure 4.** *SVR9* and *SVR9-like* genes in *Arabidopsis.* A, Gene models of three *Arabidopsis*
682 genes coding for putative prokaryotic type IF3 proteins. Gene models were drawn as in Fig.
683 2A. Numbers of nucleotides of each exon excluding the lengths of UTRs were labeled on top
684 of each exon. B, Transient expression of SVR9-GFP, SVR9L1-GFP or GFP alone in wild type
685 *Arabidopsis* leaf protoplasts. A single representative protoplast was shown for each
686 transformation. Merged images from GFP and chlorophyll channels were shown in the
687 "Merge" lane. BF, bright field. Bars, 10 μm. C-D, Tissue expression patterns of *SVR9*.
688 Illustrated are GUS staining of 6-day-old (C) and 2-week-old (D) transgenic plants expressing
689 the transcriptional fusion of P_{SVR9}*:GUS*.
690

691 **Figure 5.** Genetic interactions between *var2*, *svr9-2* and *svr9l1-1*. A-D, The first true leaves
692 of representative 2-week-old WT, *var2-5* and *var2-4* (A), *svr9l1-1*, *svr9-2* and *svr9l1-1 svr9-2*
693 double mutant (B), *svr9l1-1 var2-5* double mutant, *svr9-2 var2-5* double mutant, and *svr9l1-1*
694 *svr9-2 var2-5* triple mutant (C), *svr9l1-1 var2-4* double mutant, *svr9-2 var2-4* double mutant
695 and *svr9l1-1 svr9-2 var2-4* triple mutant (D). E-H, Quantitative comparison of leaf
696 variegation based on the frequency distributions of the pixel intensity values of leaf images
697 shown in A-D.
698
699 **Figure 6.** Functional redundancy of *SVR9* and *SVR9L1*. A, Overall phenotypes of 2-week-old
700 WT, *svr9l1-1*, *svr9-1* and *svr9-1 svr9l1-1*/+. B, Seed settings of WT and *svr9-1 svr9l1-1*/+.
701 Developing siliques of WT and *svr9-1 svr9l1-1*/+ at the same stage were dissected and
702 photographed with a stereoscope. White arrows indicated the abolished embryos in *svr9-1*
703 *svr9l1-1*/+ silique. Bars, 2.5 mm. C, Overall phenotypes of 2-week-old WT, *svr9-1* and a
704 representative line overexpressing *SVR9L1* in the *svr9-1* background.
705
706 **Figure 7.** Leaf development phenotypes in *svr9-1*, *svr9l1-1* and *svr9-1 svr9l1-1*/+ mutants. A,
707 Photographs of the third true leaves of representative 2-week-old WT, *svr9l1-1*, *svr9-1* and
708 *svr9-1 svr9l1-1*/+. Bars, 1 mm. B-D, Quantitative comparisons of leaf shapes in WT, *svr9l1-1*,
709 *svr9-1* and *svr9-1 svr9l1-1*/+ based on the leaf dissection index ($perimeter^2/4\pi \times$leaf area) (B),
710 the number of teeth/leaf perimeter (C), and the tooth area/leaf area (D). All measurements
711 were performed on the third true leaves of 2-week-old plants. 10 leaves of each genotype
712 were included in the statistical analysis. Data were presented as mean ± s.d. ***: $p<0.001$. E,
713 Cross sections of the basal part of the first true leaves of 10-day-old WT, *svr9-1* and *svr9-1*
714 *svr9l1-1*/+. Bars, 50 μm. F, Common cotyledon vein patterns observed in WT, *svr9-1* and
715 *svr9-1 svr9l1-1*/+.
716
717 **Figure 8.** Repression of chloroplast translation alters *DR5:GUS* expression pattern. Images of
718 three representative cotyledons were shown for each genotype or treatment. A-C, Comparison
719 of *DR5:GUS* expression patterns in WT, *svr9-1* and *svr9-1 svr9l1-1*/+ backgrounds.
720 *DR5:GUS* activities were assayed in cotyledons of 6-day-old seedlings grown on solid 1/2
721 MS medium. D-F, Effects of chloroplast translation inhibitors on *DR5:GUS* expression. 6-
722 day-old wild type background *DR5:GUS* seedlings maintained in liquid 1/2 MS culture were
723 treated with 5 mM chloramphenicol, 5 mM spectinomycin or equal amounts of ethanol (mock
724 treatment) for 24 hours before assaying *DR5:GUS* activities. Bars, 1 mm.
725
726 **Figure 9.** Effects of auxin transport inhibitors on leaf margin development. A-B, WT, *svr9-1*
727 and *svr9-1 svr9l1-1*/+ were grown on solid 1/2 MS medium supplemented with 1 μM TIBA

(A), 1 μM NPA (B) or equal amounts of DMSO (mock treatment) for 14 days. The third true leaves from representative plants of each genotype were illustrated. Bars, 1 mm. C-D, Quantification of the effects of TIBA (C) and NPA (D) on leaf shape by leaf dissection index measurements. Data were presented as mean ± s.d. **:0.001<p<0.01; ***: p<0.001.

Figure 10. Genetic interactions between *cuc2* mutants and *svr9-1*, *svr9l1-1* mutants. A, Photographs of the third true leaves in representative 2-week-old WT, *cuc2-101*, *cuc2-1D*, *svr9-1*, *cuc2-101 svr9-1*, *cuc2-1D svr9-1*, *svr9-1 svr9l1-1*/+, *cuc2-101 svr9-1 svr9l1-1*/+, and *cuc2-1D svr9-1 svr9l1-1*/+. B-D, Quantitative comparisons of leaf margins of indicated genotypes. Leaf margin quantifications were based on the leaf dissection index (B), the number of teeth/leaf perimeter (C), and the tooth area/leaf area (D). All leaf shape parameters were obtained as in Fig. 7. Data were presented as mean ± s.d. ***: p<0.001.

Tables

Table 1. Quantification of cotyledon vein patterns in wild type, *svr9-1* and *svr9-1 svr9l1-1/+*. Percentages of different types of areoles were indicated in the parentheses.

Genotype	Total*	Cotyledon vein patterns					
		Zero Areole	One Areole	Two Areoles	Three Areoles	Four Areoles	Five Areoles
WT	328	N.A.	N.A.	134 (40.8%)	135 (41.2%)	59 (18%)	N.A.
svr9-1	429	N.A.	13 (3%)	164 (38.2%)	183 (42.7%)	68 (15.9%)	1 (0.2%)
svr9-1 svr9l1-1/+	317	3 (1%)	67 (21.1%)	177 (55.8%)	69 (21.8%)	1 (0.3%)	N.A.

*Total number of cotyledons examined.

748 **Literature cited**

749 Aluru MR, Bae H, Wu D, Rodermel SR (2001) The *Arabidopsis immutans* mutation affects
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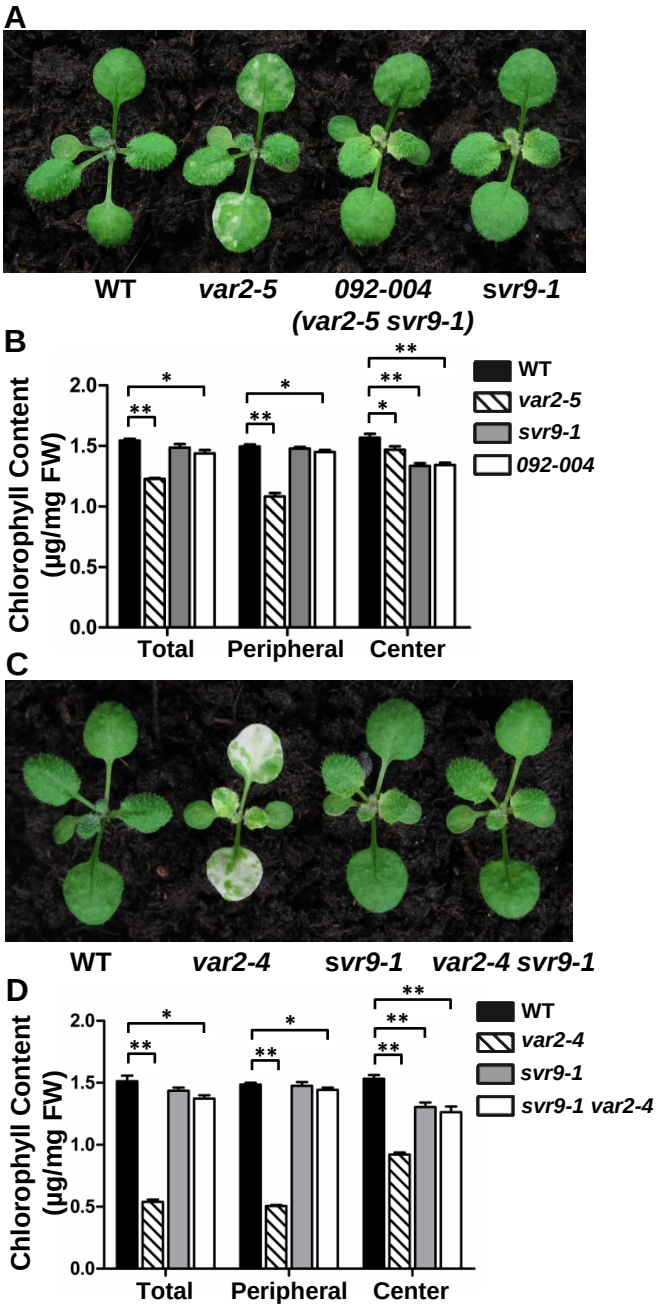

Figure 1. Suppression of *var2* leaf variegation by *svr9-1*. A, Representative 15-day-old wild type (WT), *var2-5*, *092-004* (*var2-5 svr9-1*), and *svr9-1* plants. C, Representative 15-day-old WT, *var2-4*, *svr9-1*, and *var2-4 svr9-1* double mutant. B and D, Chlorophyll contents of indicated tissues in plants shown in A and C, respectively. Total: entire rosettes excluding the cotyledons; Peripheral: the first two true leaves; Center: rosettes excluding the cotyledons and the first two true leaves. Chlorophyll measurements were normalized on a fresh tissue weight basis. Data were presented as mean ± standard deviation (s.d.) of three biological replicates. *: $0.01<p<0.05$; **: $p<0.01$.

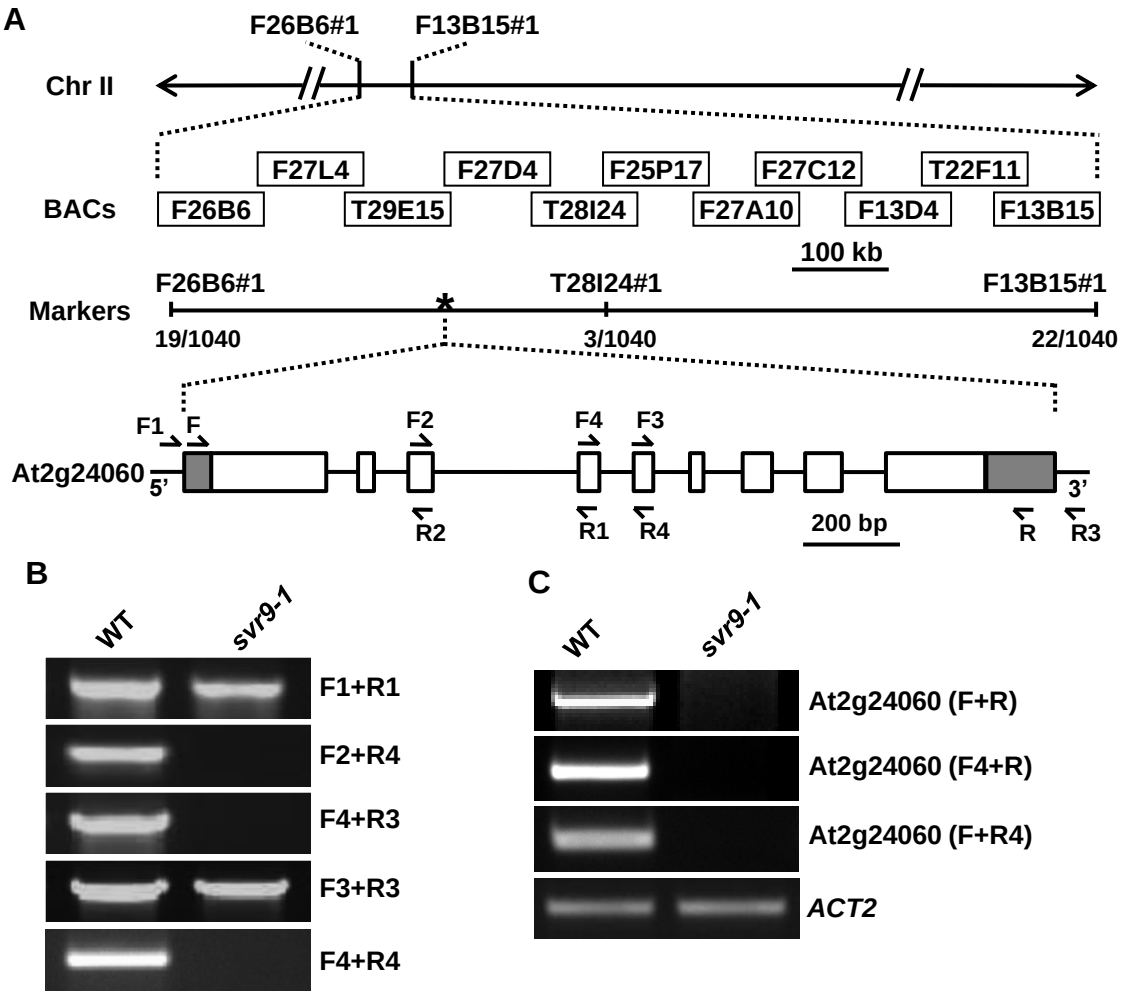

Figure 2. Cloning of *SVR9*. A, Schematic representation of the positional cloning procedure used to identify *svr9-1* mutation. Primers of the molecular markers designed in this study were listed in Supplemental Table S2. A total of 570 F2 plants (1140 chromosomes) were used in fine mapping. Numbers of recombinants were marked under each molecular marker. The asterisk indicated the position of *SVR9* (At2g24060). In the gene model, boxes represent exons and lines represent introns. 5' and 3' untranslated regions (UTRs) were shaded. Arrows represent the positions of the primers used in B and C. B, PCR analyses using various primer combinations indicated that At2g24060 genomic regions were disrupted in *svr9-1*. C, Semi-quantitative RT-PCR analyses of the accumulations of *SVR9* transcripts in wild type and *svr9-1* using indicated primer combinations. Expression of *ACT2* was used as an internal control.

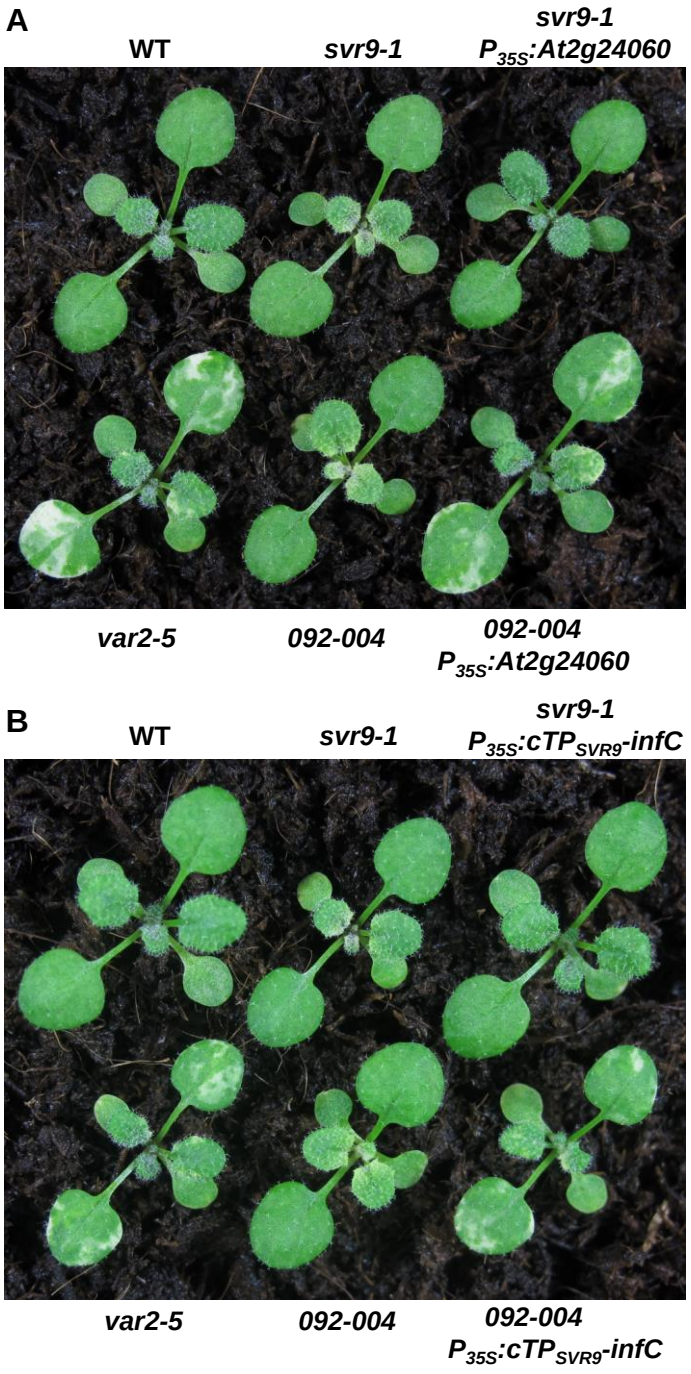

Figure 3. Complementation of *svr9-1* and *092-004*. A, Leaf color phenotypes of two-week-old WT, *svr9-1*, *var2-5*, *092-004* and representative lines overexpressing At2g24060 in *svr9-1* or *092-004* background. B, Leaf color phenotypes of two-week-old WT, *svr9-1*, *var2-5*, *092-004* and representative lines overexpressing CTP_{SVR9}-infC in *svr9-1* or *092-004* background.

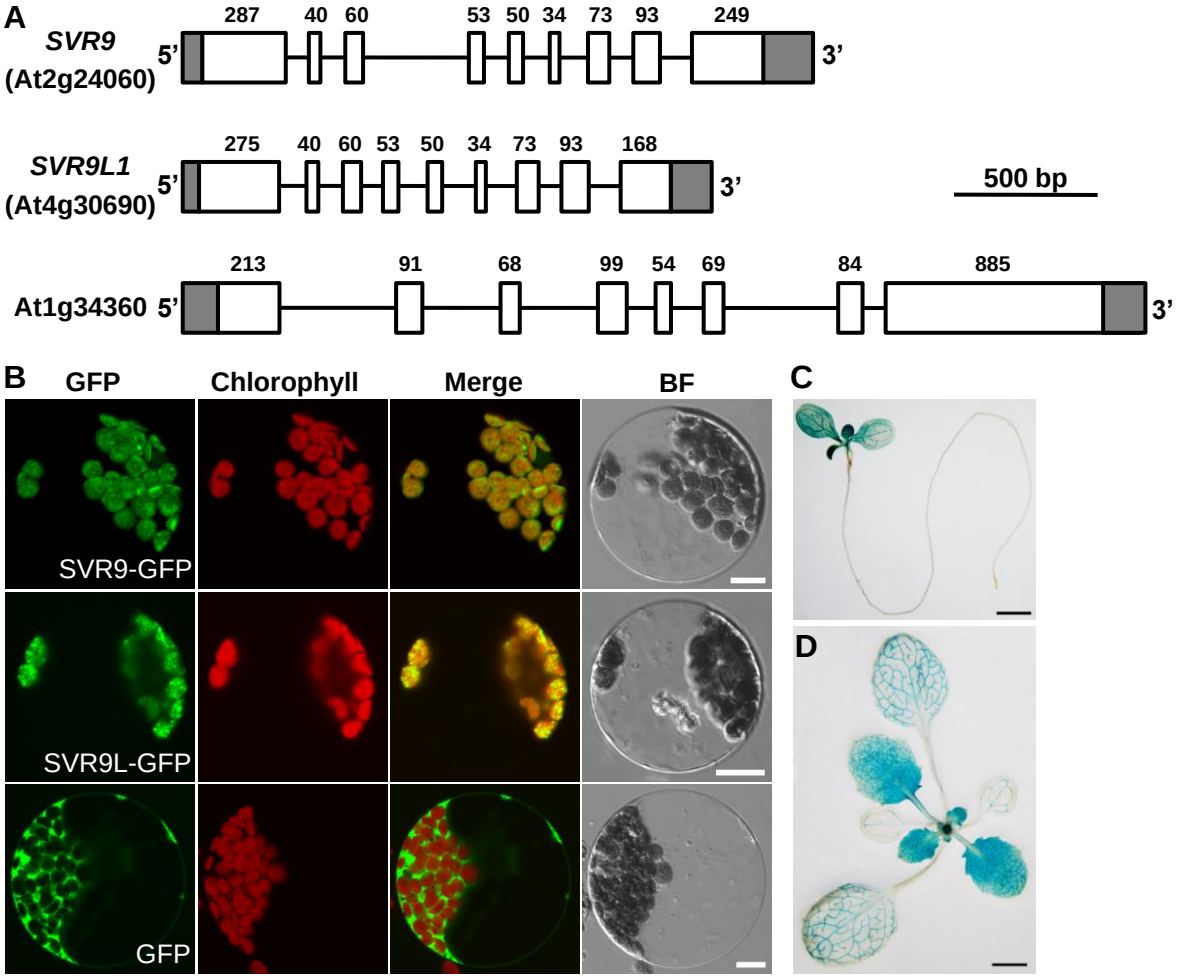

Figure 4. *SVR9* and *SVR9-like* genes in *Arabidopsis.* A, Gene models of three *Arabidopsis* genes coding for putative prokaryotic type IF3 proteins. Gene models were drawn as in Fig. 2A. Numbers of nucleotides of each exon excluding the lengths of UTRs were labeled on top of each exon. B, Transient expression of SVR9-GFP, SVR9L1-GFP or GFP alone in wild type *Arabidopsis* leaf protoplasts. A single representative protoplast was shown for each transformation. Merged images from GFP and chlorophyll channels were shown in the "Merge" lane. BF, bright field. Bars, 10 μm. C-D, Tissue expression patterns of *SVR9*. Illustrated are GUS staining of 6-day-old (C) and 2-week-old (D) transgenic plants expressing the transcriptional fusion of P_{SVR9}:*GUS*.

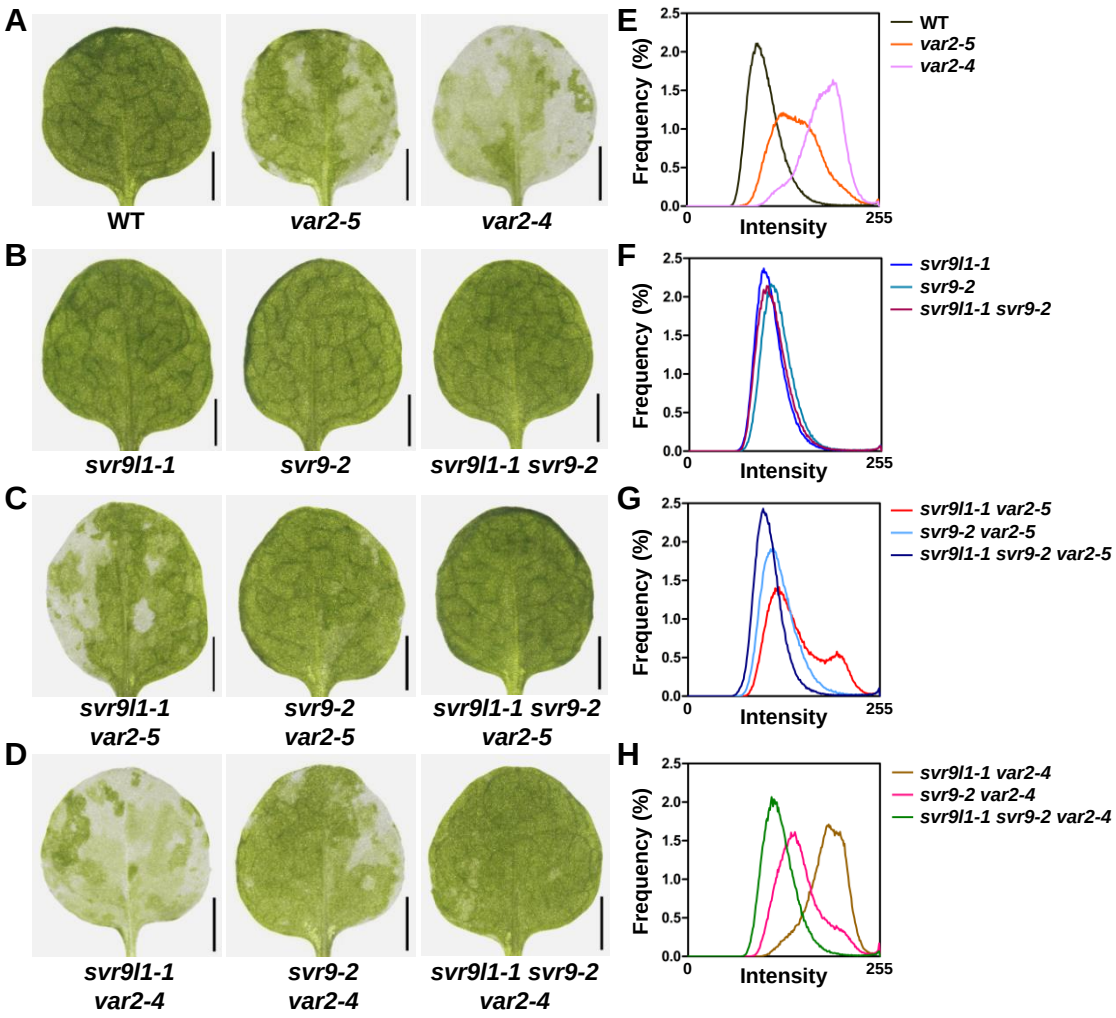

Figure 5. Genetic interactions between *var2*, *svr9-2* and *svr9l1-1*. A-D, The first true leaves of representative 2-week-old WT, *var2-5* and *var2-4* (A), *svr9l1-1*, *svr9-2* and *svr9l1-1 svr9-2* double mutant (B), *svr9l1-1 var2-5* double mutant, *svr9-2 var2-5* double mutant, and *svr9l1-1 svr9-2 var2-5* triple mutant (C), *svr9l1-1 var2-4* double mutant, *svr9-2 var2-4* double mutant and *svr9l1-1 svr9-2 var2-4* triple mutant (D). E-H, Quantitative comparison of leaf variegation based on the frequency distributions of the pixel intensity values of leaf images shown in A-D.

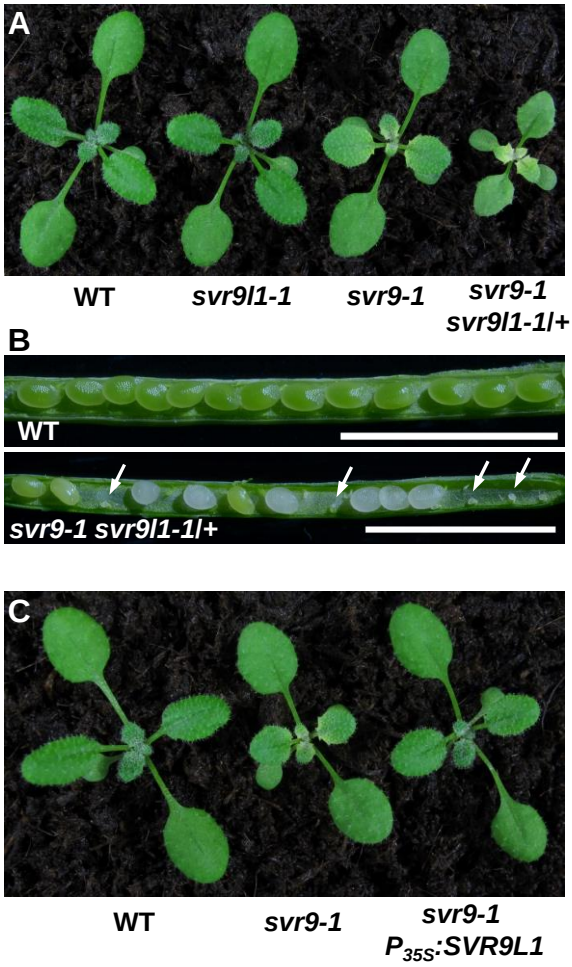

Figure 6. Functional redundancy of *SVR9* and *SVR9L1*. A, Overall phenotypes of 2-week-old WT, *svr9l1-1*, *svr9-1* and *svr9-1 svr9l1-1*/+. B, Seed settings of WT and *svr9-1 svr9l1-1*/+. Developing siliques of WT and *svr9-1 svr9l1-1*/+ at the same stage were dissected and photographed with a stereoscope. White arrows indicated the abolished embryos in *svr9-1 svr9l1-1*/+ silique. Bars, 2.5 mm. C, Overall phenotypes of 2-week-old WT, *svr9-1* and a representative line overexpressing *SVR9L1* in the *svr9-1* background.

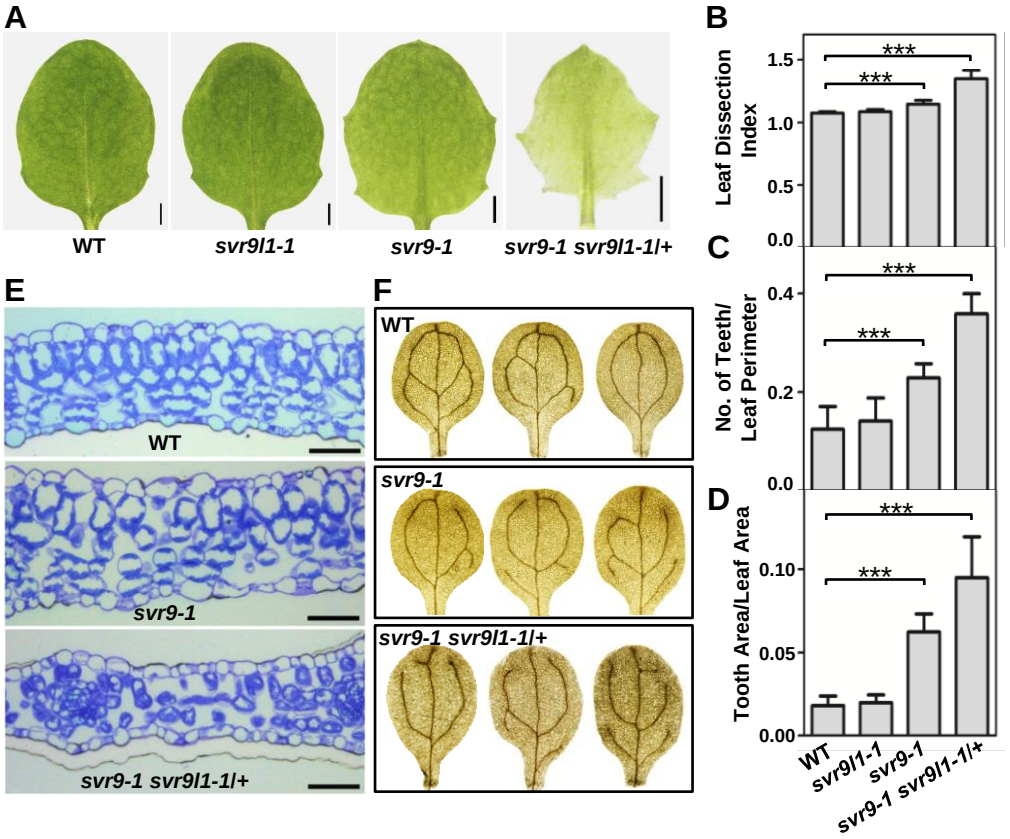

Figure 7. Leaf development phenotypes in *svr9-1*, *svr9l1-1* and *svr9-1 svr9l1-1*/+ mutants. A, Photographs of the third true leaves of representative 2-week-old WT, *svr9l1-1*, *svr9-1* and *svr9-1 svr9l1-1*/+. Bars, 1 mm. B-D, Quantitative comparisons of leaf shapes in WT, *svr9l1-1*, *svr9-1* and *svr9-1 svr9l1-1*/+ based on the leaf dissection index (perimeter^2/4π×leaf area) (B), the number of teeth/leaf perimeter (C), and the tooth area/leaf area (D). All measurements were performed on the third true leaves of 2-week-old plants. 10 leaves of each genotype were included in the statistical analysis. Data were presented as mean ± s.d. ***: p<0.001. E, Cross sections of the basal part of the first true leaves of 10-day-old WT, *svr9-1* and *svr9-1 svr9l1-1*/+. Bars, 50 μm. F, Common cotyledon vein patterns observed in WT, *svr9-1* and *svr9-1 svr9l1-1*/+.

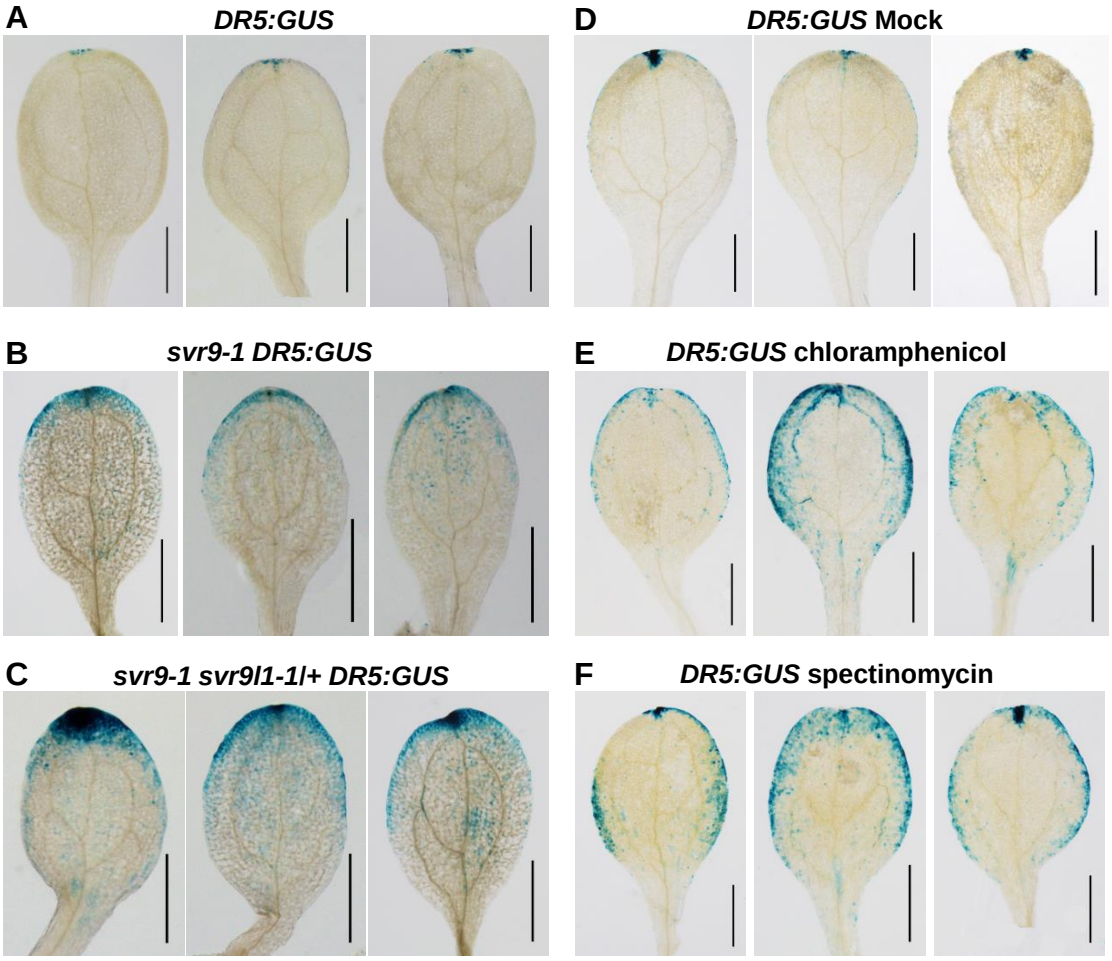

Figure 8. Repression of chloroplast translation alters *DR5:GUS* expression pattern. Images of three representative cotyledons were shown for each genotype or treatment. A-C, Comparison of *DR5:GUS* expression patterns in WT, *svr9-1* and *svr9-1 svr9l1-1*/+ backgrounds. *DR5:GUS* activities were assayed in cotyledons of 6-day-old seedlings grown on solid 1/2 MS medium. D-F, Effects of chloroplast translation inhibitors on *DR5:GUS* expression. 6-day-old wild type background *DR5:GUS* seedlings maintained in liquid 1/2 MS culture were treated with 5 mM chloramphenicol, 5 mM spectinomycin or equal amounts of ethanol (mock treatment) for 24 hours before assaying *DR5:GUS* activities. Bars, 1 mm.

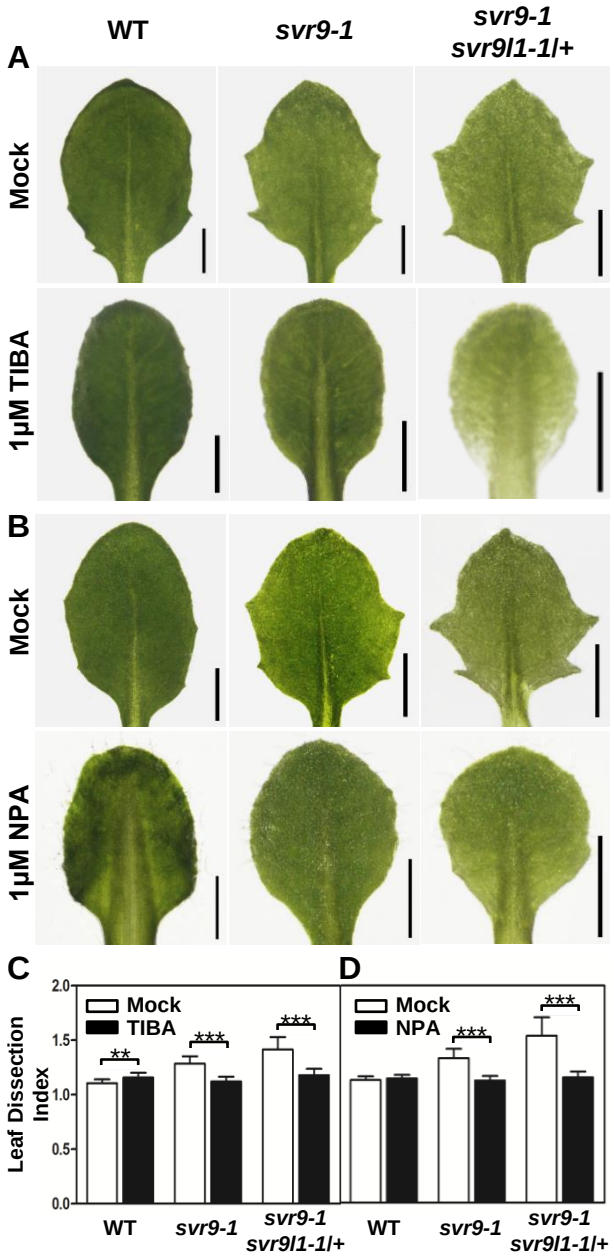

Figure 9. Effects of auxin transport inhibitors on leaf margin development. A-B, WT, *svr9-1* and *svr9-1 svr9l1-1*/+ were grown on solid 1/2 MS medium supplemented with 1 μM TIBA (A), 1 μM NPA (B) or equal amounts of DMSO (mock treatment) for 14 days. The third true leaves from representative plants of each genotype were illustrated. Bars, 1 mm. C-D, Quantification of the effects of TIBA (C) and NPA (D) on leaf shape by leaf dissection index measurements. Data were presented as mean ± s.d. **:0.001<p<0.01; ***: p<0.001.

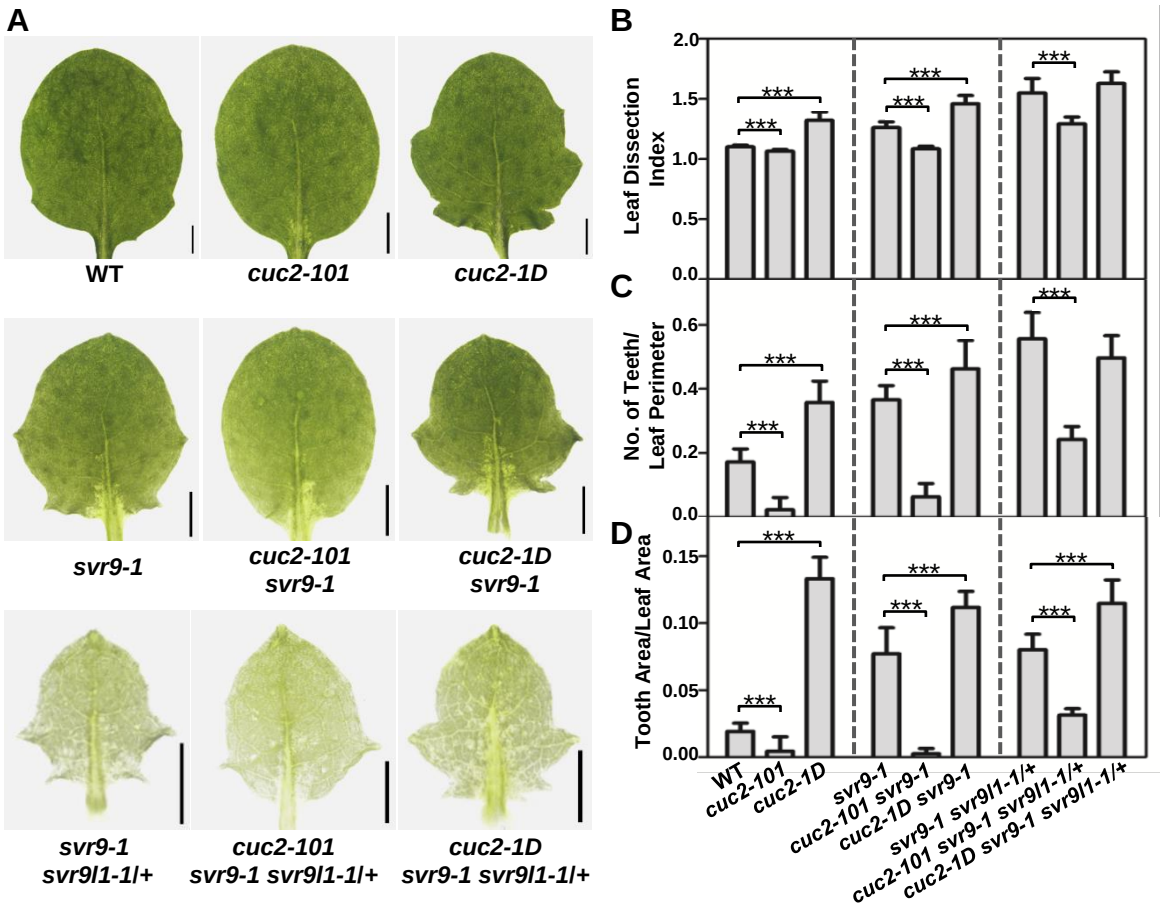

Figure 10. Genetic interactions between *cuc2* mutants and *svr9-1*, *svr9l1-1* mutants. A, Photographs of the third true leaves in representative 2-week-old WT, *cuc2-101*, *cuc2-1D*, *svr9-1*, *cuc2-101 svr9-1*, *cuc2-1D svr9-1*, *svr9-1 svr9l1-1*/+, *cuc2-101 svr9-1 svr9l1-1*/+, and *cuc2-1D svr9-1 svr9l1-1*/+. B-D, Quantitative comparisons of leaf margins of indicated genotypes. Leaf margin quantifications were based on the leaf dissection index (B), the number of teeth/leaf perimeter (C), and the tooth area/leaf area (D). All leaf shape parameters were obtained as in Fig. 7. Data were presented as mean ± s.d. ***: p<0.001.

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