

Article type : Research Article

**Host-induced gene silencing of an important pathogenicity factor
PsCPK1 in *Puccinia striiformis* f. sp. *tritici* enhances resistance of wheat
to stripe rust**

Tuo Qi, Xiaoguo Zhu, Chenlong Tan, Peng Liu, Jia Guo, Zhensheng Kang, Jun
Guo*

State Key Laboratory of Crop Stress Biology for Arid Areas, College of Plant
Protection, Northwest A&F University, Yangling 712100, Shaanxi, P. R. China

Tuo Qi: 2014060057@nwsuaf.edu.cn

Xiaoguo Zhu: 877812121@qq.com

Chenglong Tan: 935750186@qq.com

Peng Liu: 114732842@qq.com

Jia Guo: 450630983@qq.com

Zhensheng Kang: kangzs@nwsuaf.edu.cn

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/pbi.12829

This article is protected by copyright. All rights reserved.

Accepted Article

*Author for correspondence: Jun Guo, Tel: 0081-29-87082439, E-mail:
guojunwgq@nwafu.edu.cn.

Running title: HIGS of *PsCPK1* enhances wheat resistance to *Pst*

Key words: host-induced gene silencing, *PsCPK1*, wheat, *Puccinia striiformis*
f. sp. *tritici*, virulence, RNAi

SUMMARY

Rust fungi are devastating plant pathogens and cause a large economic impact on wheat production worldwide. To overcome this rapidly loss of varieties resistance, we generated stable transgenic wheat plants expressing short interfering RNAs (siRNAs) targeting potentially vital genes of *Puccinia striiformis* f. sp. *tritici* (*Pst*). Protein kinase A (PKA) has been proved to play important roles in regulating the virulence of phytopathogenic fungi. *PsCPK1*, a PKA catalytic subunit gene from *Pst*, is highly induced at the early infection stage of *Pst*. The instantaneous silencing of *PsCPK1* by *barley stripe mosaic virus* (BSMV)-mediated host-induced gene silencing (HIGS) results in a significant reduction in the length of infection hyphae and disease phenotype. These results indicate that *PsCPK1* is an important pathogenicity factor by regulating *Pst* growth and development. Two transgenic lines expressing the RNA interference (RNAi) construct in a normally susceptible wheat cultivar

This article is protected by copyright. All rights reserved.

Accepted Article

displayed high levels of stable and consistent resistance to *Pst* throughout the T₃ to T₄ generations. The presence of the interfering RNAs in transgenic wheat plants was confirmed by northern blotting, and these RNAs were found to efficiently down-regulate *PsCPK1* expression in wheat. The present study addresses important aspects for the development of fungal-derived resistance through the expression of silencing constructs in host plants as a powerful strategy to control cereal rust diseases.

INTRODUCTION

Wheat stripe rust, caused by *Puccinia striiformis* f. sp. *tritici* (*Pst*), is one of the most serious diseases of wheat (*Triticum aestivum*) worldwide (Chen *et al.*, 2013). This disease can result in more than 90% yield losses in a field (<http://striperust.wsu.edu>). The most economical, effective, and environmentally friendly way of controlling this disease is to breed and use wheat varieties. However, most race-specific host resistance genes have transient protection, probably due to the rapid evolution of new virulent rust fungal isolates (Fisher *et al.*, 2012). Therefore, new feasible methods must be uncovered to protect wheat crops from rust fungi.

RNA interference (RNAi) is firstly discovered in *Caenorhabditis elegans* (Fire *et al.*, 1998) and it can trigger potent and specific interference of pathogenic processes through exogenous double-stranded RNA (dsRNA). For several insects and nematodes, silencing has been confirmed by merely feeding the dsRNA of the target genes (Hannon, 2002; Huvenne and Smaghe, 2010).

With the identification of small RNAs (siRNAs), the mechanism of gene silencing of essential cellular functions was established (Castel and Martienssen, 2013; Vaucheret and Fagard, 2001). RNAi has been proved to be a powerful tool to reveal functions of target genes in organisms (Hellens *et al.*, 2005) and offers an effective strategy to enhance resistance in crop plants. The accumulation of RNAi molecules in barley targeting fungal transcripts affects the development of *Blumeria graminis* f. sp. *hordei* in barley (Nowara *et al.*, 2010). Transient silencing genes encoding MAP kinases, calcineurin B (PtCNB) and cyclophilin (PtCYC1), lead to increased resistance in wheat against rust diseases (Panwar *et al.*, 2013a). Expressing inverted repeat fragments of Cellulose Synthase (CES1) genes of *Bremia lactucae* in transgenic plants resulted in attenuated pathogenicity and growth of *B. lactucae* (Govindarajulu *et al.*, 2015). Three RNAi constructs derived from *Chs3b* were identified as the most effective RNAi constructs for enhancing resistance to *Fusarium* pathogens *in planta* (Cheng *et al.*, 2015).

As a biotrophic parasite, *Pst* infects the host mainly from urediospores, which germinate within 3 h at a low-temperature after deposition on the leaf surface (Hassebrauk and Schroeder, 1964). Germ tubes grow perpendicular to the long axis of epidermal cells of the leaf until they encounter a stoma. At 6-8 hours post-inoculation (hpi), an appressorium forms above the stoma and subsequently a substomatal vesicle forms within the stomatal cavity. At 12-18 hpi the primary infection hypha and haustorial mother cells emerge. Haustorial mother cells (HMCs), which have a thick, multilayered wall, invaginate the host cell plasma membrane with a slender neck to form the haustorium (Kang *et al.*,

2003). Haustoria withdraw nutrients from host cells through the extrahaustorial matrix (Voegelé and Mendgen, 2003). The primary infection hyphae branch and produce a number of HMCs and haustoria from 24 to 144 hpi, developing into the fungal mycelium within the host tissue. From 6 to 8 days after infection symptoms of chlorosis will be observed, whereas sporulation commences after approximately 12 to 14 days under favorable conditions.

The cyclic adenosine monophosphate-protein kinase A (cAMP-PKA) signaling pathway is well conserved across eukaryotes and has been proved to participate in virulence, morphogenesis, and development in diverse fungi (Bahn and Sundstrom, 2001; D'Souza and Heitman, 2001; Fuller and Rhodes, 2012). In *Saccharomyces cerevisiae*, cAMP-dependent protein kinase has a vital role in controlling proliferation, stress resistance, metabolism, and the availability of nutrients (Thevelein and De Winder, 1999; Toda *et al.*, 1987). The catalytic subunits of PKA including *Tpk1-3* have different functions. *Tpk2* has a unique role in activation of pseudohyphal growth. In contrast, *Tpk1* and *Tpk3* repress filamentation (Pan and Heitman, 1999; Robertson and Fink, 1998). Further studies revealed that *Tpk2* has a negative role in regulating genes for iron uptake whereas it has a positive role in regulating genes for water homeostasis and trehalose degradation (D'Souza *et al.*, 2001). In *Magnaporthe oryzae*, *CPKA* was shown to inhibit appressorium formation and the responsiveness of germinating conidia to exogenous cAMP (Mitchell and Dean, 1995). The maintenance of pathogenicity in *cpkA* mutants on wounded plants implies an additional role of *CPKA* that may be essential for appressorial penetration (Xu *et al.*, 1997). In *Fusarium graminearum*, *CPK1* is

responsible for hyphal growth, differentiation, and pathogenesis (Hu *et al.*, 2014). In *Ustilago maydis*, two genes *adr1* and *uka1* are found to encode catalytic subunits of PKA. The *adr1* is the major PKA catalytic subunit gene and required for pathogenicity, whereas *uka1* has almost no influence on pathogenicity (Dürrenberger *et al.*, 1998). The availability of rust fungus genomic resources (Duplessis *et al.*, 2011; Xu *et al.*, 2011; Zheng *et al.*, 2013) accelerate the prediction of a great number of genes and the research on functional genomics of rust fungi. However, due to the lack of functional genomics tools for rust fungi, less is known about biological functions of these genes.

In this study, we characterized a gene encoding the catalytic subunit of PKA, designated *PsCPK1* in *Pst*. The results showed that knockdown of *PsCPK1* leads to decreased virulence of *Pst*. The hairpin silencing constructs of *PsCPK1* expressed in wheat plants are sufficient to suppress disease development of *Pst*, indicating durable resistance at the genetic level against *Pst* infection.

RESULTS

***Pst* contains two PKA catalytic subunit genes**

A BLAST search using *adr1* and *uka1* of *U. maydis* (Dürrenberger *et al.*, 1998) as queries revealed the genome of *Pst* contains two PKA catalytic subunit genes, *PSTG06781* and *PSTG11839*, that were named *PsCPK1* and *PsCPK2*,

Accepted Article

respectively (Zheng *et al.*, 2013). In this study, sequence analysis indicated that *PsCPK1* has an open reading frame (ORF) of 1443 bp, encoding a putative protein composed of 480 amino acids with a molecular weight of 55.69 kD and an isoelectric point (pI) of 6.72. A multi-sequence alignment with seven CPK proteins of different organisms in NCBI database revealed that *PsCPK1* is 80% and 57.3% identical to *CPK1* from *Puccinia graminis* f. sp. *tritici* (*Pgt*) and *Puccinia triticina* (*Pt*), respectively, and contain almost all the conserved domains of the PKA catalytic subunit. Compared to the *PsCPK1* protein, *PsCPK2* shares 30.57% similarity and 40.54% identity at the nucleotide sequence level. Phylogenetic analysis indicated that the homologs of *CPK* in *Pst* separate into two distinct groups, class I and class II (Figure 1). *PsCPK1* in class I is orthologous to *adr1* of *U. maydis*, *cpkA* of *M. oryzae*, and yeast *TPK2*, whereas *PsCPK2* in class II is orthologous to *cpk2* of *M. oryzae*, and *uka1* of *U. maydis*. These results indicate that *CPK* is highly conserved in other filamentous fungi.

PsCPK1* is highly expressed at the early infection stage of *Pst

To investigate whether *PsCPK1* is involved in *Pst* infection, quantitative RT-PCR (qRT-PCR) was used to test *PsCPK1* transcript levels in different *Pst* infection stages. The transcript level of *PsCPK1* was gradually induced as early as 6 hpi and at 18 hpi attained the maximum level of 11.7-fold compared with that in the control. Then the transcript level returned to the original level at 24 to 48 hpi (Figure 2). Its expression down-regulated between 72 to 264 hpi

Accepted Article

and was barely detected at the sporulation stage (216-264 hpi). Our results indicate that the transcription of *PsCPK1* is induced during the infection stage.

***PsCPK1* partially complements the *M. oryzae cpkA* mutant**

The PsCPK1 protein shares 80% similarity and 66% identity with CPK1 of *M.*

No lesions were found on leaves sprayed with water or conidia of DF51 (Figure 3c). Our data show that *PsCPK1* can partially complement the DF51 in appressorium formation and plant infection.

Transient silencing of *PsCPK1* significantly reduces pathogenicity of *Pst*

BSMV-mediated HIGS was adopted to silence *PsCPK1* in *Pst*. Two specific fragments of *PsCPK1* were designed for silencing this gene (Figure S1). Ten days after infection with BSMV, wheat seedlings inoculated with sterile FES buffer developed normal leaves (Figure 4a). Under the same conditions, photo-bleaching was observed in BSMV:TaPDS-as infected plants (Figure 4a), indicating that *TaPDS* was successfully and specifically silenced. All wheat seedlings inoculated with BSMV:γ (control), BSMV:PsCPK1-1as and BSMV:PsCPK1-2as showed mild chlorotic mosaic symptoms on the third leaf (Figure 4a), suggesting that the BSMV-HIGS system functioned well. In comparison with the control plants, wheat leaves infected with BSMV:PsCPK1-1as and BSMV:PsCPK1-2as displayed a susceptible phenotype with similar viral infection symptoms (Figure 4b). However, the number of uredia in BSMV:PsCPK1-1as and BSMV:PsCPK1-2as infected leaves was significantly lower than that in the control (Figure 4b). Compared with control plants, the ratio of pustules on leaves infected with BSMV:PsCPK1-1as and BSMV:PsCPK1-2as was reduced by 49% and 54%, respectively (Figure 4c). To test silencing efficiency of BSMV-HIGS, qRT-PCR was used to assay the relative transcript level of *PsCPK1* at 24, 48, 120 hpi with the virulent CYR32

isolate. Compared with control plants, the transcript level in the BSMV:PsCPK1-1as infected leaves was reduced by 74%, 85% and 58%, respectively (Figure 4d). Similarly, in BSMV:PsCPK1-2as infected leaves, the transcript level of *PsCPK1* was reduced by 75%, 81% and 62%, respectively (Figure 4d).

The detailed histological changes in HIGS-silenced plants inoculated with *Pst* CYR32 were microscopically examined (Figure S2 a-f). In BSMV:PsCPK1-1as and BSMV:PsCPK1-2as inoculated leaves, the hyphal lengths exhibited significant reduction relative to that in the control plants at 48 hpi and 120 hpi (Figure S2 g). The infection areas in the silenced plants also decreased at 120 hpi (Figure S2 h). However, the numbers of haustorial mother cells showed no significant difference at 48 hpi (Figure S2 i).

Molecular analysis of transgenic wheat plants

To efficiently generate specific siRNAs directed against the *PsCPK1* gene in transgenic wheat plants, the coding region of *PsCPK1* (1443 bp) was cloned into the plasmid pMCG161 and the cloned fragments as inverted repeats under control of the maize ubiquitin promoter (*Ubi1*) to generate the dsRNA (Figure 5a). The resultant construct was then bombarded into wheat cultivar (cv.) Xinong1376 (XN1376). In the greenhouse experiments, ten transgenic lines (L1, L2, L5, L9, L10, L11, L12, L14, L17 and L18) (Table S1) were tested and the disease severity scale was evaluated by the standard described in Table S2 (Line and Qayoum, 1992). Among these ten lines, the transgenic L12

Accepted Article

and L18 lines were selected for further study (Figure S3). Integration of the transgene in the two lines L12 and L18 carrying the resultant constructs was confirmed by genomic PCR with primers Bar-F/R, UBI1-F/R and TG-CPK1-F/R (Table S3, Figure 5a). To further verify that these two lines were independently derived and transgenic, Southern blotting was performed with genomic DNA from the two lines digested with *Bam*HI and *Xho*I. The results showed that each line contained one copy and exhibited a different banding pattern (Figure 5b). To confirm whether HIGS efficiently works to protect transgenic wheat plants from *Pst* infection, northern blotting was performed to detect *PsCPK1* dsRNA expression in L12 and L18. The results show that L12 and L18 produce the small interfering RNAs (siRNAs, ~21 nt) (Figure 5c, Figure S4).

Expression of *PsCPK1* small RNA in wheat confers durable resistance to *Pst*

Pst resistance was evaluated in the greenhouse. Average reduction of disease severity in L12 and L18 was 59% and 46%, respectively (Figure S3).

Phenotypically, the second leaves of 14-day-old L12 and L18 plants were challenged with the virulent CYR32 isolate (Figure 6a). All of the infected leaves could be separated into three classes (Table S2) and the ratio of diseased plants in the 1-3 grade was 69% and 77% in L12 and L18 (Figure 6b). To determine the transcript levels of *PsCPK1* at different infection stages of *Pst* on XN1376, we performed qRT-PCR assay (Figure S5). According to the results, we isolated total RNA from silenced leaves after 48, 72, 120, 168

Accepted Article

hpi with *Pst* urediospores to test silencing efficiency of *PsCPK1*. Relative expression analysis of these samples compared with transgenic controls revealed specific reductions in transcript levels of the *CPK1* gene in *Pst*. The transcript levels of the *PsCPK1* showed an approximate 42% to 64% down-regulation in L12 and 36% to 50% in L18 compared with controls (Figure 6c), suggesting *PsCPK1* suppression had an approximate relevance to increased resistance against *Pst* in L12 and L18. Additionally, a series of experiments provided evidence of the effectiveness and stability of the transgenes. To determine the ratio of sporulating to non-sporulating uredia, a quantitative assay was made within a defined surface area of the leaves at 16 days after *Pst* infection, which is consistent with the rust disease phenotype. The ratio of sporulating to non-sporulating uredia was significantly reduced by about 80% in transgenic lines. Fungal biomass in the infected leaves was also measured. Total genomic DNA was isolated from wheat leaves infected with *Pst*, and the relative levels of *PsEF1* and *TaEF1* were quantified by Q-PCR (Figure S6). Compared with controls, fungal biomass was significantly reduced by about 76% in L12 and 72% in L18 at 7 dpi, respectively (Figure 6e). Together these results indicate that in transgenic wheat plants *PsCPK1* expression is efficiently down-regulated and *Pst* growth is impaired and delayed.

Histological and molecular changes of *Pst* growth in transgenic plants

To further test the possible function of the *PsCPK1* in *Pst*-wheat interaction, we performed microscopic assessment of *Pst* development in foliar tissue. The infection sites were chosen randomly from the second infected leaves of L12 and L18 plants and transgenic control plants at 48 hpi and 120 hpi and analyzed by fluorescence microscopy. At 48 hpi, fungal penetration and expansion abilities were not different, and the mycelial morphology appeared normal compared with that in the transgenic control lines (Figure 7a-c). Mycelial morphology appeared normal (Figure 7d-f), whereas at 168 hpi both the length of infection hyphae (IH) (Figure 7g) and area of infection unit was significantly reduced (Figure 7h). There were no significant differences on the number of haustorial mother cells at 48 and 168 hpi (data not shown). To further understand the relationship between transgenic wheat and controls, we assayed the transcript profiles of a few selected genes after infection with *Pst*. The transcript levels of *PsRAS2*, upstream of the cAMP-PKA pathway in *Pst*, were down-regulated in transgenic lines compared with the control (Figure 8), but *PsRAS1* was only down-regulated at 48 hpi. We also subsequently monitored the relative expression of several other subunit genes of the PKA complex (*PsCPK2* and *PsRPK*) in transgenic plants (Figure 8). Interestingly, both were down-regulated at 168 hpi, but showed no obvious changes at 48 hpi (Figure 8). *PsPrf1*, a transcription factor functioning after phosphorylation by *PsCPK1*, decreased significantly in the L12 and L18 lines at 48 hpi and 168 hpi. Therefore, our data indicate that *PsCPK1* most likely plays an important in

the virulence of *Pst* by participating in fungal development and growth, and silencing of the *PsCPK1* results in virulence penalty of *Pst* in transgenic plants.

DISCUSSION

To compensate for the limited amount of germplasm resources and rapid loss of resistance to fungal diseases, several new approaches have emerged in recent years. Many previous studies have experimentally validated that RNAi is a promising approach for durable control of pathogenic fungi (Ghag *et al.*, 2014; Chen *et al.*, 2016; Cheng *et al.*, 2015; Nowara *et al.*, 2010). In the rust fungi, the RNAi approach has been developed through BSMV (Yin *et al.*, 2011). The *Agrobacterium tumefaciens*-mediated transient RNAi assay has been developed in wheat to target *P. triticina* pathogenicity genes (Panwar *et al.*, 2013a). But, complete resistance has rarely been achieved in wheat, and it is therefore significant that transgenic resistance to *Pst* was identified to the fourth generation in our study. To our knowledge this is the first reported transgenic wheat with durable disease resistance against *Pst*.

A few studies have proved that expression of dsRNA targeted at fungal genes in wheat leads to increased resistance against phytopathogenic fungi (Chen *et al.*, 2016; Cheng *et al.*, 2015). Usually true transgenic lines in hexaploid wheat are available until the T₃ generation (Lee *et al.*, 2012) and then we generated T₄ generation for further investigation. By analyzing the T₄ generation, we confirmed by northern blot analysis that transgene-derived

siRNAs accumulate in the resistant transgenic plants. Moreover, those plants had the strongest reduction in mRNA and corresponding decrease in susceptibility. Southern blot analysis indicated that each resistant transgenic line contains a single copy derived from independent transformations. Unlike the resistance of germplasm resources, the reduction of the disease phenotype does not depend on race-specific resistance (Johnson, 1988) largely because of the high conservation of *PsCPK1* in most isolates (data not shown), including the popular isolates CYR32 and V26 (Huang *et al.*, 2014; Ren *et al.*, 2015; Yin *et al.*, 2009). Moreover, we found that transient silencing by BSMV-HIGS is maintained for only 2 weeks, as indicated by Yin *et al.* (2011) and Miller *et al.* (2012), whereas the transgenic plants show a slow but continuous weakening of *Pst* silencing. Our results suggest that transgenic wheat has the potential to reduce the severity of stripe rust of wheat.

To further understand the mechanism after silencing the *PsCPK1*, we investigated the transcript levels of some important genes in the cAMP-PKA pathway and the other subunit genes of the PKA complex. Ras proteins function at upstream of mitogen-activated protein kinase (MAPK) or cAMP-PKA pathway (Bluhm *et al.*, 2007; Park *et al.*, 2006). In *Pst*, *PsRas2* is required for *Pst* pathogenicity but not for *PsRas1* (Cheng *et al.*, 2016). *PsRas2* and *PsRas1* were significantly reduced after *PsCPK1* silencing, perhaps through the feedback loop of the cAMP (Figure S7). The genes *PsCPK2* and *PsRPK*, encoding another catalytic subunit and the regulatory subunit of the PKA complex, were down-regulated at 168 hpi. Interaction of the catalytic subunits of PKA with the regulatory subunit of PKA was activated by cAMP.

Silencing of *PsCPK1* may destroy the structure of the PKA complex. Since *Prf1* is required for cell fusion and filamentous growth (Hartmann *et al.*, 1999) and the activity of *Prf1* is controlled by PKA phosphorylation (Zarnack *et al.*, 2008), a domino effect may appear in the transgenic plants after silencing of the *PsCPK1*. Together with these results, *PsCPK1* plays important roles in the cAMP-PKA pathway and silencing the *PsCPK1* will disrupt the chain of cAMP-PKA pathway after infection by *Pst* (Figure S7).

In the process of the application of RNAi methods in organisms the off-target effects maybe occur when the siRNA is partially complementary to one or more cellular mRNAs except the target (de Souza, 2014; Jackson *et al.*, 2003). Despite the high conservation of *CPK1* in different fungi, the *PsCPK1* is sequence-specific among its homologues in pathogenic fungi at the nucleotide level. Additionally, the homologous *PsCPK1* gene in wheat shares up to 30% identity at the nucleotide sequence level within the coding regions. The possibility of off-target effects was analyzed by Si-Fi software and no effective hits were found (Table S4). These off-targets effects can show no obvious difference on the growth of wheat (Figure S8). Furthermore, the off-target effect could be more pronounced in rust fungi, which might have resulted in the disease suppression.

HIGS has been proved to be a novel tool to reveal gene function in obligate biotrophic fungus (Panwar *et al.*, 2013a; Yin *et al.*, 2011) and offers the potential of disease control (Nowara *et al.*, 2010). In this study wheat plants expressing the RNAi constructs showed strong and genetically stable

Accepted Article

resistance to *Pst* in the fourth generation. Our results indicated that *PsCPK1* is an excellent target to generate durable genetic resistance against *Pst* and provides a potential reservoir of novel resistance resources of wheat against rust fungi. But the application of this material in agricultural production obviously requires more field trials. With the rapid development of various sequencing technologies, quantitative proteomics and RNA-seq analysis may be used in this material to understand the mechanism of the cAMP-mediated signal transduction pathway. The improved understanding will guide the development of genetic breeding, which could contribute to environmentally sustainable agriculture.

EXPERIMENTAL PROCEDURES

Biological materials, growth condition, fungal inoculation

The wheat cv. Suwon11 (Su11) and the *Pst* virulent isolate CYR32 were used in this study (Guo *et al.*, 2011). *Pst* isolate CYR32 was maintained and propagated on susceptible wheat cv. Mingxian169. The germplasm used for gene transformation was XN1376, a high yielding and early maturing winter wheat variety. Plant cultivation and fungal inoculation following the procedures and conditions previously described (Kang *et al.*, 2002).

Plasmid construction and plant transformation

To generate pMCG161-CPK1, the PCR was performed to obtain the product with the primers PsCPK1-F and PsCPK1-R (Table S3). The amplified fragment was subsequently cloned into the *Ascl* and *AvrII* and *AsiSI* and *SpeI* sites of plasmid pMCG161. Then the CPK1+-intro-CPK1-gene from T-simple (TaKaRa, Tokyo, Japan) was cut with *Ascl* and *AvrII*, and ligated into the pMCG161 with *Ascl* and *AvrII*.

Sequence alignments and polymorphism analysis

One cDNA clone encoding *PsCPK1* (GenBank accession #KY346510) was designed with special primers CPK1-F/R (Table S3) according to the sequence from the cDNA library during wheat-*Pst* interaction (Ma *et al.*, 2009). *PsCPK1* sequence was firstly analyzed using BLAST search and ORF Finder, then the conserved domain of PsCPK1 was detected with InterProScan and ScanProsite (Guo *et al.*, 2013). Multiple sequence alignment was implemented with DNAMAN6.0 (Lynnon BioSoft, USA) and CLUSTALX2.0 (Chenna *et al.*, 2003). The phylogenetic tree was constructed with the Mega 6.0 software (Tamura *et al.*, 2013).

RNA extraction, cDNA synthesis and qRT-PCR

To evaluate the transcript levels of *PsCPK1* in response to *Pst* infection, wheat leaves were sampled at 0, 6, 12, 18, 24, 36, 48, 72, 120, 144, 216, 264 hpi according to previous microscopic observations of wheat-*Pst* interaction (Wang *et al.*, 2007). To determine the efficiency of BSMV-mediated HIGS, the fourth leaves of *PsCPK1*-knockdown plants were sampled at 24, 48, 120 hpi with *Pst*. Total RNA was isolated with the Trizol reagent (Invitrogen, Carlsbad, CA, USA) and the first strand cDNA was synthesized for qRT-PCR. qRT-PCR was performed with a 7500 Real-Time PCR System (Applied Biosystems, Foster City, CA, USA) and PCR conditions were used as previously described (Liu *et al.*, 2012). The *Pst* translation elongation factor 1 (*PsEF1*) gene was used as the internal reference for normalization (Guo *et al.*, 2011). The transcript levels of *PsCPK1* and other genes in this study were assayed by the comparative $2^{-\Delta\Delta CT}$ method (Livak and Schmittgen, 2001).

Complementation of the *M. oryzae cpkA* mutant with *PsCPK1*

To perform complementation assays, the full-length of the *PsCPK1* gene was obtained with primers CPK1-CM-F and CPK1-CM-R (Table S3). Then the *PsCPK1* gene was co-transformed with the vector pFL2 into *S. cerevisiae* strain XK1-25 to achieve plasmid pMoPsCPK1. The plasmid pMoPsCPK1 was transformed into protoplasts of the *M. oryzae cpkA* mutant DF51. To confirm the *PsCPK1* gene integrated into the *M. oryzae* genome, we isolated the resultant transformants and verified them by PCR with primers CPK1-CM-F

and CPK1-CM-R. The assays for appressorium formation and plant infection were performed as previously described (Guo *et al.*, 2011).

BSMV-mediated *PsCPK1* gene silencing

To further determine the role of *PsCPK1* during *Pst* infection, the *PsCPK1* gene was silenced with the BSMV-HIGS system. To make sure the specificity for *PsCPK1* silencing, a 224-bp fragment of *PsCPK1* in the 5'UTR named *PsCPK1-1as* and a 296-bp fragment of *PsCPK1* in the 3'UTR named *PsCPK1-2as* were cloned with primers Higs-CPK1-1as-F, Higs-CPK1-1as-R, Higs-CPK1-2as-F and Higs-CPK1-2as-R (Table S3) and inserted into the virus plasmid. The BSMV RNAs were prepared *in vitro* from linearized plasmid γ -TaPDSas, γ -*PsCPK1-1as*, γ -*PsCPK1-2as*, γ , α , β using the Message T7 *in vitro* transcription kit (Ambion, Austin, TX, USA). The wheat leaves for inoculation with BSMV according to the procedures as previously described (Guo *et al.*, 2013; Scofield *et al.*, 2005). After inoculation with BSMV at the second leaf stage, wheat seedlings were maintained in a growth chamber at 23 ± 2 °C and examined for symptoms. In all experiments, the recombinant virus BSMV:TaPDSas was applied as a positive control. When the photo-bleaching phenotype was observed, the fourth leaves of *PsCPK1* silencing group were inoculated with urediospores of *Pst* isolate CYR32. The resistant or susceptible phenotypes were visible at 15 dpi.

Histological observations of *Pst* growth and wheat response

The wheat leaves inoculated with BSMV at 24, 48 and 120 hpi and transgenic wheat leaves at 48 and 168 hpi were collected for histological observation as previously described (Guo *et al.*, 2013). Stained leaf segments were fixed and cleared in ethanol/acetic acid (1:1 v/v). Auto-fluorescence of attacked mesophyll cells was observed as a necrotic death area with the Olympus BX-51 microscope. Infection sites and lengths of infection hyphae were measured under the blue light excitation. Fifty infection sites were examined on each randomly selected leaf segment per treatment for measurement of fungal structures.

Northern blotting analysis

Northern blotting was used to detect accumulation of small RNAs as described previously (Zheng *et al.*, 2007). The total RNA from 12-day-old seedlings was extracted using Trizol reagent (Invitrogen, Carlsbad, CA, USA). The poly(ethylene glycol) enrichment method was used to obtain small RNA (Zheng *et al.*, 2007). The random priming method was used to label the probe for detection of mRNA transcripts. The primers NB-CPK1-F and NB-CPK1-R used to obtain the probe are listed in Table S3.

Southern blotting analysis

A CTAB-based method was used to isolate total genomic DNA from 12-day-old seedlings (Hormaza, 2002). Aliquots (20 µg) of genomic DNA were digested overnight at 37°C with *Bam*HI and *Xho*I, fractionated in 0.8% (w/v) agarose gel and blotted onto a nylon membrane (BioTrace, Gelman Sciences) according to the protocols previously described (Sambrook and Russell, 1989). The 302-bp cDNA fragment was obtained by RT-PCR with primer SB-CPK1-F/R (Table S3) and then used as the probe. The membrane was hybridized with the probe labelled with [α -³²P] dCTP. Hybridization, post hybridization washes and signal detection were carried out as described for northern blotting (Sambrook and Russell, 1989), but the hybridization temperature was adjusted to 65°C.

Statistical analysis

Statistical testing was performed with the statistical software version package of IBM SPSS Statistics 21 (IBM SPSS Statistics, IBM Corporation, Armonk, New York). The data were tested by Student's test ($p < 0.05$ or $p < 0.01$).

ACKNOWLEDGEMENTS

This work was financially supported by the National Natural Science Foundation of China (31620103913 and 31371889) and the National Basic Research Program of China (2013CB127700). The authors declare no conflict of interest.

REFERENCES

- Bahn, Y. and Sundstrom, P. (2001) *CAP1*, an adenylate cyclase-associated protein gene, regulates bud-hypha transitions, filamentous growth, and cyclic AMP levels and is required for virulence of *Candida albicans*. *J. Bacteriol.* **183**, 3211-3223.
- Bluhm, B.H., Zhao, X., Flaherty, J.E., Xu, J. and Dunkle, L.D. (2007) RAS2 regulates growth and pathogenesis in *Fusarium graminearum*. *Mol. Plant Microbe Interact.* **20**, 627-636.

- Chen, W., Wellings, C., Chen, X., Kang, Z. and Liu, T. (2013) Wheat stripe (yellow) rust caused by *Puccinia striiformis* f. sp. *tritici*. *Mol. Plant Pathol.* **15**, 433-446.
- Cheng, W., Song, X., Li, H., Cao, L., Sun, K., Qiu, X., Xu, Y., Yang, P., Huang, T., Zhang, J., Qu, B. and Liao, Y. (2015) Host-induced gene silencing of an essential chitin synthase gene confers durable resistance to *Fusarium* head blight and seedling blight in wheat. *Plant Biotechnol. J.* **13**, 1335-1345.
- Cheng, Y., Wang, W., Yao, J., Huang, L., Voegelé, R.T., Wang, X.J. and Kang, Z.S. (2016) Two distinct *Ras* genes from *Puccinia striiformis* exhibit differential roles in rust pathogenicity and cell death. *Environ. Microbiol.* **18**, 3910-3922.
- Chenna, R., Sugawara, H., Koike, T., Lopez, R., Gibson, T.J., Higgins, D.G. and Thompson, J.D. (2003) Multiple sequence alignment with the Clustal series of programs. *Nucleic Acids Res.* **31**, 3497-3500.
- de Souza, N. (2014) Genetics: more specific CRISPR editing. *Nat. Methods* **11**, 712.
- D'Souza, C.A. and Heitman, J. (2001) Conserved cAMP signaling cascades regulate fungal development and virulence. *Fems. Microbiol. Rev.* **25**, 349-364.
- D'Souza, C.A., Alspaugh, J.A., Yue C., Harashima, T., Cox, G.M., Perfect, J.R. and Heitman, J. (2001) Cyclic AMP-dependent protein kinase controls

virulence of the fungal pathogen *Cryptococcus neoformans*. *Mol. Cell. Biol.* **21**, 3179-3191.

Duplessis, S., Cuomo, C.A., Lin, Y., Aerts, A., Tisserant, E., Veneault-Fourrey, C., Joly, D.L., Hacquard, S., Amselem, J. and Cantarel, B.L. (2011) Obligate biotrophy features unraveled by the genomic analysis of rust fungi. *Proc Natl Acad Sci USA.* **108**, 9166-9171.

Dürrenberger, F., Wong, K. and Kronstad, J.W. (1998) Identification of a cAMP-dependent protein kinase catalytic subunit required for virulence and morphogenesis in *Ustilago maydis*. *Proc Natl Acad Sci USA.* **95**, 5684-5689.

Fire, A., Xu, S., Montgomery, M.K., Kostas, S.A., Driver, S.E. and Mello, C.C. (1998) Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature* **391**, 806-811.

Fisher, M.C., Henk, D.A., Briggs, C.J., Brownstein, J.S., Madoff, L.C., McCraw, S.L. and Gurr, S.J. (2012) Emerging fungal threats to animal, plant and ecosystem health. *Nature* **484**, 186-194.

Fuller, K.K. and Rhodes, J.C. (2012) Protein kinase A and fungal virulence. *Virulence* **3**, 109-121.

Ghag, S.B., Shekhawat, U.K.S. and Ganapathi, T.R. (2014) Host-induced post-transcriptional hairpin RNA-mediated gene silencing of vital fungal genes confers efficient resistance against *Fusarium* wilt in banana. *Plant Biotechnol. J.* **12**, 541-553.

- Govindarajulu, M., Epstein, L., Wroblewski, T. and Michelmore, R.W. (2015)
Host-induced gene silencing inhibits the biotrophic pathogen causing
downy mildew of lettuce. *Plant Biotechnol. J.* **13**, 875-883.
- Guo, J., Bai, P., Yang, Q., Liu, F., Wang, X., Huang, L. and Kang, Z. (2013)
Wheat zinc finger protein TaLSD1, a negative regulator of programmed
cell death, is involved in wheat resistance against stripe rust fungus. *Plant
Physiol. Biochem.* **71**, 164-172.
- Guo, J., Dai, X., Xu, J., Wang, Y., Bai, P., Liu, F., Duan, Y., Zhang, H., Huang,
L. and Kang, Z. (2011) Molecular characterization of a Fus3/Kss1 type
MAPK from *Puccinia striiformis* f. sp. *tritici*, PsMAPK1. *PLoS ONE* **6**,
e21895.
- Hannon, G.J. (2002) RNA interference. *Nature* **418**, 244-251.
- Hartmann, H.A., Krüger, J., Lottspeich, F. and Kahmann, R. (1999)
Environmental signals controlling sexual development of the corn smut
fungus *Ustilago maydis* through the transcriptional regulator Prf1. *Plant
Cell* **11**, 1293-1305.
- Hassebrauk, K. and Schroeder, J. (1964) Studies on the germination of yellow
rust urediospores. In *Proceedings of the First European and
Mediterranean Cereal Rusts Conference*, pp.12-18. Cambridge.
- Hellens, R.P., Allan, A.C., Friel, E.N., Bolitho, K., Grafton, K., Templeton, M.D.,
Karunairetnam, S., Gleave, A.P. and Laing, W.A. (2005) Transient

expression vectors for functional genomics, quantification of promoter activity and RNA silencing in plants. *Plant Methods* **1**, 13.

- Hormaza, J.I. (2002) Molecular characterization and similarity relationships among apricot (*Prunus armeniaca* L.) genotypes using simple sequence repeats. *Theor. Appl. Genet.* **104**, 321-328.
- Hu, S., Zhou, X., Gu, X., Cao, S., Wang, C. and Xu, J. (2014) The cAMP-PKA pathway regulates growth, sexual and asexual differentiation, and pathogenesis in *Fusarium graminearum*. *Mol. Plant Microbe Interact.* **27**, 557-566.
- Huang, Q., Li, X., Chen, W.Q., Xiang, Z.P., Zhong, S.F., Chang, Z.J., Zhang, M., Zhang, H.Y., Tan, F.Q. and Ren, Z.L. (2014) Genetic mapping of a putative *Thinopyrum intermedium*-derived stripe rust resistance gene on wheat chromosome 1B. *Theor. Appl. Genet.* **127**, 843-853.
- Huvenne, H. and Smagghe, G. (2010) Mechanisms of dsRNA uptake in insects and potential of RNAi for pest control: a review. *J. Insect Physiol.* **56**, 227-235.
- Jackson, A.L., Bartz, S.R., Schelter, J., Kobayashi, S.V., Burchard, J., Mao, M., Li, B., Cavet, G. and Linsley, P.S. (2003) Expression profiling reveals off-target gene regulation by RNAi. *Nat. Biotechnol.* **21**, 635-637.
- Johnson, R. (1988) Durable resistance to yellow (stripe) rust in wheat and its implications in plant breeding. In *Breeding Strategies for Resistance to the Rusts of Wheat*, pp. 63-75. CIMMYT, Mexico D.F., Mexico.

- Kang, Z., Huang, L. and Buchenauer, H. (2002) Ultrastructural changes and localization of lignin and callose in compatible and incompatible interactions between wheat and *Puccinia striiformis*. *J. Plant Dis. Protect.* **109**, 25-37.
- Kang, Z.S., Wang, Y., Huang, L.L., Wei, G.R. and Zhao, J. (2003) Histology and ultrastructure of incompatible combination between *Puccinia striiformis* and wheat cultivars with resistance of low reaction type. *Scientia Agricultura Sinica* **36**, 1026-1031.
- Lee, W., Hammond-Kosack, K.E. and Kanyuka, K. (2012) Barley stripe mosaic virus-mediated tools for investigating gene function in cereal plants and their pathogens: virus-induced gene silencing, host-mediated gene silencing, and virus-mediated overexpression of heterologous protein. *Plant Physiol.* **160**, 582-590.
- Line, R.F. and Qayoum, A. (1992) Virulence, aggressiveness, evolution and distribution of races of *Puccinia striiformis* (the cause of stripe rust of wheat) in North America, pp. 1968-87. Technical Bulletin (USA).
- Liu, F., Guo, J., Bai, P., Duan, Y., Wang, X., Cheng, Y., Feng, H., Huang, L. and Kang, Z. (2001) Wheat TaRab7 GTPase is part of the signaling pathway in responses to stripe rust and abiotic stimuli. *PLoS ONE* **7**, e37146.

- Livak, K.J. and Schmittgen, T.D. (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* **25**, 402-408.
- Ma, J., Huang, X., Wang, X., Chen, X., Qu, Z., Huang, L. and Kang, Z. (2009) Identification of expressed genes during compatible interaction between stripe rust (*Puccinia striiformis*) and wheat using a cDNA library. *BMC Genomics* **10**, 586.
- Miller, S.C., Miyata, K., Brown, S.J. and Tomoyasu, Y. (2012) Dissecting systemic RNA interference in the red flour beetle *Tribolium castaneum*: parameters affecting the efficiency of RNAi. *PLoS ONE* **7**, e47431.
- Mitchell, T.K. and Dean, R.A. (1995) The cAMP-dependent protein kinase catalytic subunit is required for appressorium formation and pathogenesis by the rice blast pathogen *Magnaporthe grisea*. *Plant Cell* **7**, 1869-1878.
- Nowara, D., Gay, A., Lacomme, C., Shaw, J., Ridout, C., Douchkov, D., Hensel, G., Kumlehn, J. and Schweizer, P. (2010) HIGS: host-induced gene silencing in the obligate biotrophic fungal pathogen *Blumeria graminis*. *Plant Cell* **22**, 3130-3141.
- Pan, X. and Heitman, J. (1999) Cyclic AMP-dependent protein kinase regulates pseudohyphal differentiation in *Saccharomyces cerevisiae*. *Mol. Cell Biol.* **19**, 4874-4887.
- Panwar, V., McCallum, B. and Bakkeren, G. (2013a) Endogenous silencing of *Puccinia triticina* pathogenicity genes through in *planta*-expressed

sequences leads to the suppression of rust diseases on wheat. *Plant J.* **73**, 521-532.

- Panwar, V., McCallum, B. and Bakkeren, G. (2013b) Host-induced gene silencing of wheat leaf rust fungus *Puccinia triticina* pathogenicity genes mediated by the Barley stripe mosaic virus. *Plant Mol. Biol.* **81**, 595-608.
- Park, G., Xue, C., Zhao, X., Kim, Y., Orbach, M. and Xu, J. (2006) Multiple upstream signals converge on the adaptor protein Mst50 in *Magnaporthe grisea*. *Plant Cell* **18**, 2822-2835.
- Ren, Y., Li, S., Xia, X., Zhou, Q., He, Y., Wei, Y., Zheng, Y. and He, Z. (2015) Molecular mapping of a recessive stripe rust resistance gene *yrMY37* in Chinese wheat cultivar Mianmai 37. *Mol. Breeding* **35**, 1-9.
- Robertson, L.S. and Fink, G.R. (1998) The three yeast A kinases have specific signaling functions in pseudohyphal growth. *Proc Natl Acad Sci USA* **95**, 13783-13787.
- Sambrook, J. and Russell, D.W. (1989) *Molecular Cloning*. New York. A *Laboratory Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.
- Scofield, S.R., Huang, L., Brandt, A.S. and Gill, B.S. (2005) Development of a virus-induced gene-silencing system for hexaploid wheat and its use in functional analysis of the *Lr21*-mediated leaf rust resistance pathway. *Plant Physiol.* **138**, 2165-2173.

- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.* **30**, 2725-2729.
- Thevelein, J.M. and De Winde, J.H. (1999) Novel sensing mechanisms and targets for the cAMP-protein kinase A pathway in the yeast *Saccharomyces cerevisiae*. *Mol. Microbiol.* **33**, 904-918.
- Toda, T., Cameron, S., Sass, P., Zoller, M. and Wigler, M. (1987) Three different genes in *S. cerevisiae* encode the catalytic subunits of the cAMP-dependent protein kinase. *Cell* **50**, 277-287.
- Vaucheret, H. and Fagard, M. (2001) Transcriptional gene silencing in plants: targets, inducers and regulators. *Trends Genet.* **17**, 29-35.
- Voegelé, R.T. and Mendgen, K. (2003) Rust haustoria: nutrient uptake and beyond. *New Phytol.* **159**, 93-100.
- Wang, C., Huang, L., Buchenauer, H., Han, Q., Zhang, H. and Kang, Z. (2007) Histochemical studies on the accumulation of reactive oxygen species (O_2^- and H_2O_2) in the incompatible and compatible interaction of wheat-*Puccinia striiformis* f. sp. *tritici*. *Physiol. Mol. Plant Pathol.* **71**, 230-239.
- Xu, J., Linning, R., Fellers, J., Dickinson, M., Zhu, W., Antonov, I., Joly, D.L., Donaldson, M.E., Eilam, T. and Anikster, Y. (2011) Gene discovery in EST sequences from the wheat leaf rust fungus *Puccinia triticina* sexual spores, asexual spores and haustoria, compared to other rust and corn smut fungi. *BMC Genomics* **12**, 1.

- Xu, J., Urban, M., Sweigard, J.A. and Hamer, J.E. (1997) The *CPKA* gene of *Magnaporthe grisea* is essential for appressorial penetration. *Mol. Plant Microbe Intract.* **10**, 187-194.
- Yin, C., Chen, X., Wang, X., Han, Q., Kang, Z. and Hulbert, S.H. (2009) Generation and analysis of expression sequence tags from haustoria of the wheat stripe rust fungus *Puccinia striiformis* f. sp. *tritici*. *BMC Genomics* **10**, 1.
- Yin, C., Jurgenson, J.E. and Hulbert, S.H. (2011) Development of a host-induced RNAi system in the wheat stripe rust fungus *Puccinia striiformis* f. sp. *tritici*. *Mol. Plant Microbe Intract.* **24**, 554-561.
- Zarnack, K., Eichhorn, H., Kahmann, R. and Feldbrügge, M. (2008) Pheromone-regulated target genes respond differentially to MAPK phosphorylation of transcription factor Prf1. *Mol. Microbiol.* **69**, 1041-1053.
- Zheng, W.M., Huang, L.L., Huang, J.Q., Wang, X.J., Chen, X.M, Zhao, J., Guo, J., Zhuang, H., Qiu, C.Z., Liu, J., Liu, H.Q., Huang, X.L., Pei, G.L., Zhan, G.M., Tang, C.L., Cheng, Y.L., Liu, M.J., Zhang, J.S., Zhao, Z.T., Zhang, S.J., Han, Q.M., Han, D.J., Zhang, H.C., Zhao, J., Gao, X.N., Wang, J.F., Ni, P.X., Dong, W., Yang, L.F., Yang, H. M., Xu, J.R., Zhang, G.Y. and Kang Z.S. (2013) High genome heterozygosity and endemic genetic recombination in the wheat stripe rust fungus. *Nat. Commun.* **4**, 2673.

Zheng, X., Zhu, J., Kapoor, A. and Zhu, J.K. (2007) Role of Arabidopsis AGO6 in siRNA accumulation, DNA methylation and transcriptional gene silencing. *EMBO J.* **26**, 1691-1701.

FIGURE LEGENDS

Figure 1. Phylogenetic analysis of the catalytic subunits of fungal cAMP-dependent protein kinases. The amino acid sequences encoded by the catalytic subunits of the cAMP-dependent protein kinases from *Puccinia striiformis* f. sp. *tritici* (PSTG), *P. triticina* (PTTG), *P. graminis* f. sp. *tritici* (PGTG), *Ustilago maydis* (UM), *Cryptococcus neoformans* (CNAG), *Fusarium graminearum* (FGSG), *Magnaporthe oryzae* (MGG), *Saccharomyces cerevisiae* (SCRG), *Neurospora crassa* (NCU) and *Aspergillus nidulans* (AN) were retrieved from the NCBI. Phylogenetic analysis was carried out with the MEGA6 software by the neighbour-joining method.

Figure 2. Transcript profiles of *PsCPK1* at different *Pst* infection stages. Wheat leaves inoculated with virulent *Pst* isolate CYR32 sampled at 0, 6, 12, 18, 24, 36, 48, 72, 120, 144, 216 and 264 hours post inoculation (hpi). Relative expression of *PsCPK1* was calculated by the comparative threshold ($2^{-\Delta\Delta CT}$) method. Mean and standard deviation were calculated with data from three

independent biological replicates. Differences were assessed using Student's t-tests. Asterisks indicate $P < 0.05$ and double asterisks indicate $P < 0.01$.

Figure 3. Complementation of the *cpkA* mutant with the *PsCPK1* fusion construct. (a) Colony morphology of *Magnaporthe oryzae* strains. Colonies of the wild-type (Guy11), *cpkA* deletion mutant (DF51), and complemented strain (CM-41) grown on PDA plates for 5 dpi. (b) Appressorium formation assay. Germ tubes from the wild-type strain (Guy11) developed appressoria by 24 hpi, but no appressorium formation was observed in the *cpkA* mutant DF51. Under the same conditions, a transformant of expressing the *PsCPK1* fusion construct (CM-41) formed appressoria. Bar, 25 μ m. (c) Barley infection assay. Left to right, barley leaves were sprayed with sterile water and conidia of Guy11, DF51, or CM-41. Typical leaves were photographed at 6 dpi post inoculation.

Figure 4. Functional assessment of *PsCPK1* in *Pst* pathogenicity determined by BSMV-mediated HIGS. (a) Mild chlorotic mosaic symptoms were observed on the fourth leaves of wheat seedlings inoculated with BSMV: γ (control), BSMV:PsCPK1-1as, and BSMV:PsCPK1-2as. No change of phenotype was observed in wheat leaves mock-inoculated with FES buffer (MOCK). Photobleaching was evident on wheat leaves infected with BSMV:TaPDS-as. (b) Phenotypes of the fourth leaves of BSMV: γ (control), BSMV:PsCPK1-1as, and BSMV:PsCPK1-2as inoculated wheat plants 14 dpi with *Pst* isolate

CYR32. (c) Quantification of the uredial density in the BSMV:γ, BSMV:PsCPK1-1as, and BSMV:PsCPK1-2as inoculated wheat plants 14 dpi with *Pst* isolate CYR32. (d) Relative transcript levels of *PsCPK1* in the BSMV:γ, BSMV:PsCPK1-1as, and BSMV:PsCPK1-2as inoculated wheat plants 24, 48 and 120 hpi with *Pst* isolate CYR32. Values are expressed relative to the endogenous *Pst* reference gene *EF1*, with the empty vector (BSMV:γ) set at 1. Values represent the means ± standard error of three independent assays. Differences were assessed using Student's t-tests, and asterisks indicate $P < 0.05$.

Figure 5. Molecular analysis of the *Pst*-resistant transgenic wheat plants. (a) Diagram showing the RNAi cassette in the construct pMCG161-RNAi for wheat transformation. T₄ plants were analyzed by genomic PCR for the presence of the selectable marker gene *bar* (I), *UBI1* promoter (II) and fragment (*PsCPK1-intro*) of the RNAi cassette (III). The III image was compiled from different pictures. (b) Southern blotting analysis of the transgenic plants. Genomic DNA isolated from control plants (CK), T₄ transgenic plants (L12 and L18) and plasmid pMCG161-PsCPK1-RNAi. DNAs were digested with *Bam*HI and *Xho*I and hybridized to the hairpin fragment probe. The image was compiled from the same picture which we removed the lanes indicating other T₄ transgenic lines. (c) The expression of the small RNA in T₄ generation lines was analyzed by RNA gel blotting. 26s rRNA, loading/blotting control. RNA blots were hybridized with the corresponding target gene-specific probes.

siRNA levels in plants silenced using the pMCG161-PsCPK1-RNAi DNA vector expressing target fungal fragments with a mixture of sense plus antisense forms (L12 and L18). No signal was detected in the transgenic lines carrying empty vector (CK). Total RNA was extracted from three different plants. Lower panels, ethidium bromide-stained rRNA as gel loading controls. The arrowhead indicates an oligonucleotide marker of 21 nucleotides.

Figure 6. Expression of *PsCPK1* small RNA in wheat confers durable resistance to *Pst*. (a) Phenotypes of the second leaves of the third and the fourth generation of wheat plants at 14 dpi with *Pst* isolate CYR32. (b) The ratio of plants in disease grades. Phenotypes were scored to indicate the frequency of symptoms, as follows: I: 1-3; II: 4-6; III: 7-9. (c) Relative transcript levels of *PsCPK1* in the second leaves of the fourth generation wheat plants at 48, 72, 120 and 168 hpi with *Pst* isolate CYR32. Values are expressed relative to the endogenous *Pst* reference gene *PsEF1*, with the empty vector (CK) set at 1. Values represent the means \pm standard error of three independent samples. (d) Quantification of the uredial density in the CK, L12, and L18 inoculated wheat plants at 16 dpi. (e) Q-PCR measurement of fungal biomass. Ratio of fungal to wheat nuclear genomes using fungal *PsEF1* and wheat *TaEF1* genes, respectively, in plants treated with variants targeting fungal genes compared with controls. Genomic DNA extracted from the second leaf from three different plants at 7 dpi. Values represent the means \pm standard error of three independent samples. Differences were assessed using

Student's t-tests. Asterisks indicate $P < 0.05$ and double asterisks indicate $P < 0.01$.

Figure 7. Histological changes of *Pst* growth in transgenic plants. (a-f) A microscopic examination revealed no obvious differences in the number of hyphal branches between the control plants (CK) and the transgenic plants (L12 and L18) at 48 hpi (a-c) and 168 hpi (d-f), respectively. (g) The hyphal lengths in the transgene lines (L12 and L18) were shorter than those observed in the control plants (CK) at 168 hpi. (h) The colony sizes in the two transgenic plants (L12 and L18) were reduced compared with the sizes observed in the control plants (CK) at 168 hpi. Differences were assessed using Student's t-tests, and asterisks indicate $P < 0.05$.

Figure 8. Transcript levels of some selected genes involved in cAMP-PKA pathway of *Pst* after *Pst* infection. *PsRAS1*, GTPase Ras1; *PsRAS2*, GTPase Ras2; *PsCPK2*, catalytic subunits 2 of PKA; *PsRPK*, regulatory subunit of PKA; *PsPrf1*, HMG box transcription factor. Values represent the means \pm standard error of three independent assays. Differences were assessed using Student's t-tests, and asterisks indicate $P < 0.05$ and double asterisks indicate $P < 0.01$.

SUPPORTING INFORMATION LEGENDS

Figure S1. Amino acid sequence alignments of PsCPK1 with other fungal catalytic subunits of PKA. Amino acid identity (black boxes) and similarity (gray boxes) are shown within the protein kinase domain. The active sites are highlighted with arrows. Asterisks indicate the residues shown to be required for the association between catalytic and regulatory subunits of PKA. The ATP binding site is indicated with a filled triangle. PSTG06781, PGTG_17146, PTTG_25458, SCRG_03512, UM_04456, MGG_06368, and FGSG_07251 were downloaded from NCBI, respectively.

Figure S2. Histological observation of fungal growth in *PsCPK1*-knockdown wheat plants after inoculation with *Pst* isolate CYR32. (a-f) Fungal growth at 48 hpi (a-c) and 120 hpi (d-f) in wheat plants inoculated with BSMV:γ, BSMV:PsCPK1-1as, and BSMV:PsCPK1-2as. (g) Hyphal length, which is the average distance from the junction of the substomatal vesicle and the hypha to the tip of the hypha, was clearly decreased in *PsCPK1*-knockdown plants infected by CYR32. (h) The infection unit area at 120 hpi per infection unit was significantly decreased in *PsCPK1*-knockdown plants infected by CYR32. (i) No significant difference in the number of haustorial mother cells was found in *PsCPK1*-knockdown plants at 48 hpi. Values represent the means ± standard errors of three independent samples. Differences were assessed using Student's t-tests. Asterisks indicate $P < 0.05$. SV, substomatal vesicle; HMC, haustorial mother cell; IH, infection hypha; HB, hyphal branch; H, haustoria.

Figure S3. Bioassay of the transgenic wheat plants for *Pst* resistance. Two-week-old plants maintained in a controlled greenhouse were inoculated with CYR32. Plants were observed for symptom development at 16 dpi. Ten lines (L1, 2, 5, 9, 10, 11, 12, 14, 16, 17 and 18) were tested and three lines had obviously reduced *Pst* severity when compared to the transgenic control line. L12 and L18 were selected for detailed molecular analysis by southern blotting and small RNA northern blotting.

Figure S4. The expression of the small RNA in T₄ generation lines was analyzed by RNA gel blotting. 26s rRNA, loading/blotting control. RNA blots were hybridized with the corresponding target gene-specific probes.

Figure S5. Transcript profiles of *PsCPK1* in the leaves of wheat cultivar XN1376 infected by *Pst* isolate CYR32. The samples were collected at 0, 24, 48, 72, 120 and 168 h post-inoculation (hpi). The data were normalized to the endogenous *Pst* reference gene *PsEF1*, and error bars represent the variation among three independent replicates.

Figure S6. Standard curves generated for the absolute quantification of *Pst* (A) and wheat (B). Threshold cycles (Ct) are plotted against the initial copy number of template DNA (10^3 , 10^4 , 10^5 , 10^6 , 10^7 , 10^8 , and 10^9). Genomic

DNAs of the wheat cv. XN1376 and *Pst* were used to construct the standard curves.

Figure S7. Schematic presentation of possible HIGS mechanisms involved in PKA pathway. Fungal dsRNA, produced inside host cells, is cleaved by the plant silencing machinery using endonuclease-type DICER enzymes into siRNAs. siRNAs transfer from wheat cells into fungi cells through extrahaustorial matrix. This space acts as a nexus of communication and transport between host or pathogen cells and their respective exterior environments. Passage through the HCW, either active or passive, occurs and, once inside the fungal haustorium, the silencing molecules trigger RNAi of their mRNA targets, and may act as primers in the fungal silencing pathway, leading to the generation of systemic silencing signals. HMC, haustorium mother cell; H, haustorium; SSV, substomatal vesicle.

Figure S8. No significant difference was observed for the growth between transgenic and control wheat lines.

Table S1. Transformation pipeline and efficiency for producing transgenic lines used in this study.

Table S2. Disease severity scale of wheat stripe rust.

Table S3. Primers designed for *PsCPK1* research.

Table S4. Prediction off-target transcripts of *PsCPK1* gene.













